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Critical Review

The Effect of High-Fat Diet on Intramyocellular Lipid Content in Healthy Adults: A Systematic Review, Meta-Analysis, and Meta-Regression

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ABSTRACT

Fatty acids are stored within the muscle as intramyocellular lipids (IMCL). Some, but not all, studies indicate that following a high-fat diet (HFD), IMCL may accumulate and affect insulin sensitivity. This systematic review and meta-analysis aimed to quantify the effects of an HFD on IMCL. It also explored the potential modifying effects of HFD fat content and duration, IMCL measurement technique, physical activity status, and the associations of IMCL with insulin sensitivity. Five databases were systematically searched for studies that examined the effect of ≥ 3 d of HFD ($>35\%$ daily energy intake from fat) on IMCL content in healthy individuals. Meta-regressions were used to investigate associations of the HFD total fat content, duration, physical activity status, IMCL measurement technique, and insulin sensitivity with IMCL responses. Changes in IMCL content and insulin sensitivity (assessed by hyperinsulinemic-euglycemic clamp) are presented as standardized mean difference (SMD) using a random effects model with 95% confidence intervals (95% CIs). Nineteen studies were included in the systematic review and 16 in the meta-analysis. IMCL content increased following HFD (SMD = 0.63; 95% CI: 0.31, 0.94, $P = 0.001$). IMCL accumulation was not influenced by total fat content ($P = 0.832$) or duration ($P = 0.844$) of HFD, physical activity status ($P = 0.192$), or by the IMCL measurement technique ($P > 0.05$). Insulin sensitivity decreased following HFD (SMD = -0.34 ; 95% CI: $-0.52, -0.16$; $P = 0.003$), but this was not related to the increase in IMCL content following HFD ($P = 0.233$). Consumption of an HFD ($>35\%$ daily energy intake from fat) for ≥ 3 d significantly increases IMCL content in healthy individuals regardless of HFD total fat content and duration of physical activity status. All IMCL measurement techniques detected the increased IMCL content following HFD. The dissociation between changes in IMCL and insulin sensitivity suggests that other factors may drive HFD-induced impairments in insulin sensitivity in healthy individuals.

This trial was registered at PROSPERO as CRD42021257984.

Keywords: intramuscular triglyceride, lipid droplet, overfeeding, insulin resistance

Introduction

Current dietary guidelines generally recommend that total fat intake should not exceed 35% of total daily energy intake [1]. However, fat intake associated with a modern Western diet can exceed this recommendation [2], contributing to the rising rates of obesity and type 2 diabetes [3]. The link between the

consumption of a high-fat diet (HFD) and the development of metabolic disease is multifactorial, but ectopic storage of excess lipids within skeletal muscle appears to be an important etiologic factor [4].

In resting human skeletal muscle, plasma-derived nonesterified fatty acids (NEFA) enter the intramyocellular lipid (IMCL) pool and are first incorporated into intramuscular triacylglycerols

Abbreviations: ¹H-MRS, H¹ magnetic resonance spectroscopy; HFD, high-fat diet; IMCL, intramyocellular lipid; IMTG, intramuscular triacylglycerols; NEFA, nonesterified fatty acids; RCT, randomized controlled trial; TAG, triacylglycerol.

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(IMTGs) within lipid droplets before the IMTG-derived fatty acids are released for oxidation [5]. Following HFD, IMTG synthesis can exceed IMTG lipolysis, resulting in the expansion of the IMCL pool [6,7]. However, not all studies observe IMCL accumulation following HFD [8,9].

This discrepancy may be because of sampling errors associated with small sample sizes in individual studies and/or the methodologic variability among studies, such as the following: 1) the difference in HFD total fat content, 2) HFD duration, and/or 3) the measurement techniques used to quantify IMCL content. Transmission electron microscopy [10] and immunofluorescence microscopy can determine fiber-type-specific IMCL content in muscle biopsies [11]. Biochemical estimates of IMCL from mixed muscle biopsy samples do not reveal fiber-type-specific IMCL content and are potentially confounded by extramyocellular lipid [12], contributing to a large variability in the measurement of IMCL content across serial muscle biopsies [13]. Finally, the noninvasive ^1H magnetic resonance spectroscopy (^1H -MRS) technique also cannot determine fiber-type-specific IMCL content, but it can distinguish between intra- and extramyocellular lipids [13]. Pooling the results of individual studies is needed to resolve discrepancies between studies and to explore potential sources of heterogeneity [14].

IMCL content is negatively associated with insulin sensitivity in healthy individuals [15–17], and following HFD, IMCL accumulation and a reduction in insulin sensitivity have also been reported [18]. Together, these findings may suggest a link between IMCL and insulin sensitivity. However, in endurance-trained individuals, high IMCL content coexists with high-insulin sensitivity [4], casting doubt on the causal relationships between IMCL and insulin sensitivity. It is possible that the storage capacity or turnover of the IMCL pool in response to HFD is a means to reduce the effects of excess lipids on insulin sensitivity [7].

Therefore, the main purpose of this study was to synthesize evidence through a systematic review and meta-analysis of the effect of an HFD on IMCL content in healthy individuals. A secondary aim was to explore the potential modifying effects of HFD total fat content, HFD duration, IMCL measurement technique, and physical activity status on IMCL content. We also aimed to delineate the relationship between HFD-associated changes in IMCL content and insulin sensitivity.

Methods

This systematic review and meta-analysis was completed in accordance with PRISMA guidelines [19] and was prospectively registered with the PROSPERO database (reference: CRD42021257984).

Search strategy

A systematic search was conducted by 2 independent reviewers (JA and KJ) on 4 electronic databases: PubMed, Cochrane CENTRAL Register of Controlled Trials, as well as CINAHL and SPORTDiscus via EBSCOhost. The final search was conducted on 1 July 2023 using a combination of synonyms and relevant medical subject headings (MeSH) terms for HFD and IMCL, or triglyceride. Publication date and language restrictions were not applied. The specific keywords and full search strategy

for each database can be found in [Supplemental Table 1](#). We also manually screened the reference lists and forwarded citations of included studies, as well as relevant review articles, to identify potentially eligible studies, were also conducted.

Inclusion criteria

Studies were required to meet the following inclusion criteria: 1) they were original research studies involving healthy human participants aged 18–64 y. Older adults were excluded because they are more likely than their younger counterparts to be taking multiple prescription medications [20]; 2) the intervention should involve ≥ 3 d of high-calorie HFD or normocaloric HFD; 3) IMCL content was measured after the intervention; 4) full text was available in English; and 5) the study was a randomized controlled trial (RCT), within-subject crossover trial, or single-arm pretest–posttest trial. Studies were required to describe the composition of the dietary intervention in adequate detail, where total fat intake was $>35\%$ of the total daily energy intake. Studies were excluded if they met the following criteria: 1) participants were recruited on the basis of any medical condition; 2) the HFD intervention was delivered in conjunction with exercise or immediately after exercise; 3) the HFD intervention followed a period of low-fat dietary intake; and 4) the article has been retracted.

Study selection

Eligible studies were collected into a single list in Microsoft Excel (Microsoft Corporation). Two reviewers (JA and KJ) independently removed duplicates and reviewed the titles and abstracts to assess eligibility for inclusion. Reviewers were not blinded to the articles, as studies have shown that the summary outcome was not affected by blinding during study selection and data extraction [21]. Articles were initially excluded based on the content of the titles and abstracts, followed by a full-text review by 2 independent reviewers (JA and KJ). Conflicts were resolved through discussion or by a third reviewer (AH) if required.

Data extraction

Data were extracted independently by 2 researchers (JA and KJ) onto a project-specific data extraction form (Microsoft Excel). The data items extracted were as follows: 1) study characteristics (design, location, and diet duration), 2) participant characteristics (sample size, sex, age, body mass, BMI (in $\text{kg}\cdot\text{m}^{-2}$), participant health status, and participant training status), 3) details of the intervention (composition of the diet, control, duration, and number of participants per trial arm) and control arm (if applicable), and 4) details of the IMCL measurement techniques. Preintervention, postintervention, and change score data for the primary outcome (IMCL content) were also extracted [mean and (SD)]. For studies that reported fiber type-specific responses in IMCL content, data for type I and type II muscle fiber IMCL responses were extracted. For studies that reported the IMCL content within multiple muscles, data for IMCL responses in each muscle were extracted. Additionally, pre and postintervention data were extracted for the following secondary outcome measures: insulin sensitivity, circulating concentrations of fasting glucose, insulin, triacylglycerol (TAG), and NEFA. If SDs were not reported, we collected other relevant data that can be converted to an SD, such as SEs, 95% confidence intervals (CIs), or *P* values. When values were presented in figure

form only, the figure was digitized using graph digitizer software [22], and the means and SD/SEM were measured manually at the pixel level to the scale provided. However, for missing and unreported data or any further details, corresponding authors of studies were contacted via email on 2 occasions within a 1-mo period.

Assessment of risk of bias in included studies

Two authors (JA and AG) independently assessed the risk of bias in included studies in accordance with the recommendations of the Cochrane Handbook for Systematic Reviews of Interventions [23]. Cochrane Risk of Bias tool (RoB 2) was used to evaluate randomized trials [24], and the risk of bias in non-randomised studies - of interventions (ROBINS-I) tool was used to evaluate nonrandomized trials [25]. The risk of bias in studies that employed a crossover design was evaluated using RoB 2 for consistency. Disagreements between reviewers were resolved by consensus with a third reviewer (AH).

The risk of bias because of missing results in a meta-analysis was explored with Egger's test of the intercept and by visually inspecting a funnel plot of the treatment effects plotted against their corresponding sampling variance.

Statistical analysis

If 2 or more studies reported the same outcome, a meta-analysis of standardized mean differences (SMDs) was performed [26]. For studies that used independent groups, SMDs were calculated as the mean difference divided by the pooled baseline SD, where the mean difference is calculated as the mean pre-post change in the intervention group minus the mean pre-post change in the control group [27]. For studies that used matched groups (crossover trials and single-arm pre-post trials), SMDs were standardized using the change score SD. If the change score SD was not reported, it was estimated using the standard formula [28] and assuming a within-groups correlation of 0.7 [29–31]. From a statistical perspective, the SMD has the same meaning regardless of study design, and thus, we followed guidelines for including different study designs in the same meta-analysis [32]. Hedges' *g* correction was applied to SMDs. Interpretation of effect size was as follows: <0.20 as trivial, 0.20–0.49 as small, 0.50–0.79 as moderate, and >0.80 as large [33].

Meta-analyses were performed with a random effects model using the restricted maximum likelihood method to estimate between-study variance. CIs and test statistics were calculated via a *t*-distribution using the Hartung-Knapp-Sidik-Jonkman approach [34]. Studies were weighted according to the inverse of the sampling variance. If a meta-analysis included >1 outcome measure from the same study, effect estimates were nested within studies using a multi-level structure [35]. Statistical heterogeneity between studies was evaluated with the χ^2 test and the I^2 statistic. Thresholds for the interpretation of I^2 were in line with Cochrane recommendations: 0–40% (“might not be important”), 30%–60% (“moderate heterogeneity”), 50%–90% (“substantial heterogeneity”), and 75%–100% (“considerable heterogeneity”) [14]. The importance of the observed I^2 value was interpreted alongside its 95% CI and the *P* value from the χ^2 test [14]. Graphic Display of Heterogeneity plots were used to investigate between-study heterogeneity and identify potentially influential studies. Statistical analyses were

conducted using package metafor in R version 4.2 (R Foundation for Statistical Computing). If a meta-analysis model includes ≥ 10 effect estimates [14], sources of heterogeneity with meta-regressions were explored. Covariates were included: IMCL measurement technique, participant physical activity status, study design, HFD duration, HFD total fat content, composition of total fat intake, and participant body mass.

Results

Literature search

A total of 2383 articles were identified, which was reduced to 1955 after removing duplicates. Following the screening of titles and abstracts, 1926 studies were excluded because they did not meet the eligibility criteria, and we were unable to access the full text of 2 studies. Following the appraisal of full texts, a total of 19 studies were suitable for inclusion in the systematic review. Three studies [36–38] could not be synthesized by meta-analysis because of insufficient data; thus, 16 studies were included in the meta-analysis (Figure 1). Six studies were controlled trials, and 10 were single-arm trials. Of the 6 controlled trials, 3 were parallel-groups, and 3 were crossover-designed trials.

Study characteristics

A total of 303 participants were included in the meta-analysis with a mean age of 28 y (range 21–44 y). Individual study characteristics and their main findings are summarized in Table 1. The mean BMI at baseline was 24.5 (range 20.6–30.1); however, only 1 study included participants with a BMI >25 [9]. Regarding the techniques used to measure IMCL, 6 studies used $^1\text{H-MRS}$ [6,18,39–42], 5 studies used biochemical extraction [43–47], and 5 used microscopy, of which 1 study used transmission electron microscopy [48] and 4 studies used immunofluorescence microscopy [7–9,49]. One study used both $^1\text{H-MRS}$ and microscopy [8].

Diet composition

In all studies, the intervention group consumed the HFD, and the comparator groups consumed either a low-fat diet or a habitual normal-fat diet. Participants in 9 studies consumed hypercaloric HFD, whereas in 7 studies normocaloric HFD. Intervention groups consumed HFD with a proportion of fat between 35% and 78% of total daily energy expenditure. Across all studies, the mean total fat intake in HFD was 243 g/d (range 123–496 g/d). The HFDs provided across studies were either high in saturated fat (>10% of daily calories) [9,46], high in polyunsaturated fat ($\leq 10\%$ of daily calories) [40], and high in unsaturated fat ($\leq 30\%$ of daily calories) [47]. Intervention duration ranged from 3 to 56 d.

IMCL content

A meta-analysis comprising 22 effect estimates from 16 studies showed that IMCL content was significantly increased following HFD (SMD = 0.63; 95% CI: 0.31, 0.94, $P < 0.001$; Figure 2). This effect size corresponds to a percentage increase of $31.4\% \pm 29.2\%$. The magnitude of heterogeneity in IMCL responses to HFD was substantial between these studies ($I^2 = 81.57\%$; $P < 0.001$). Inspection of the funnel plot showed asymmetry in the distribution of the studies included in the meta-analysis, and Egger's regression test indicated evidence of

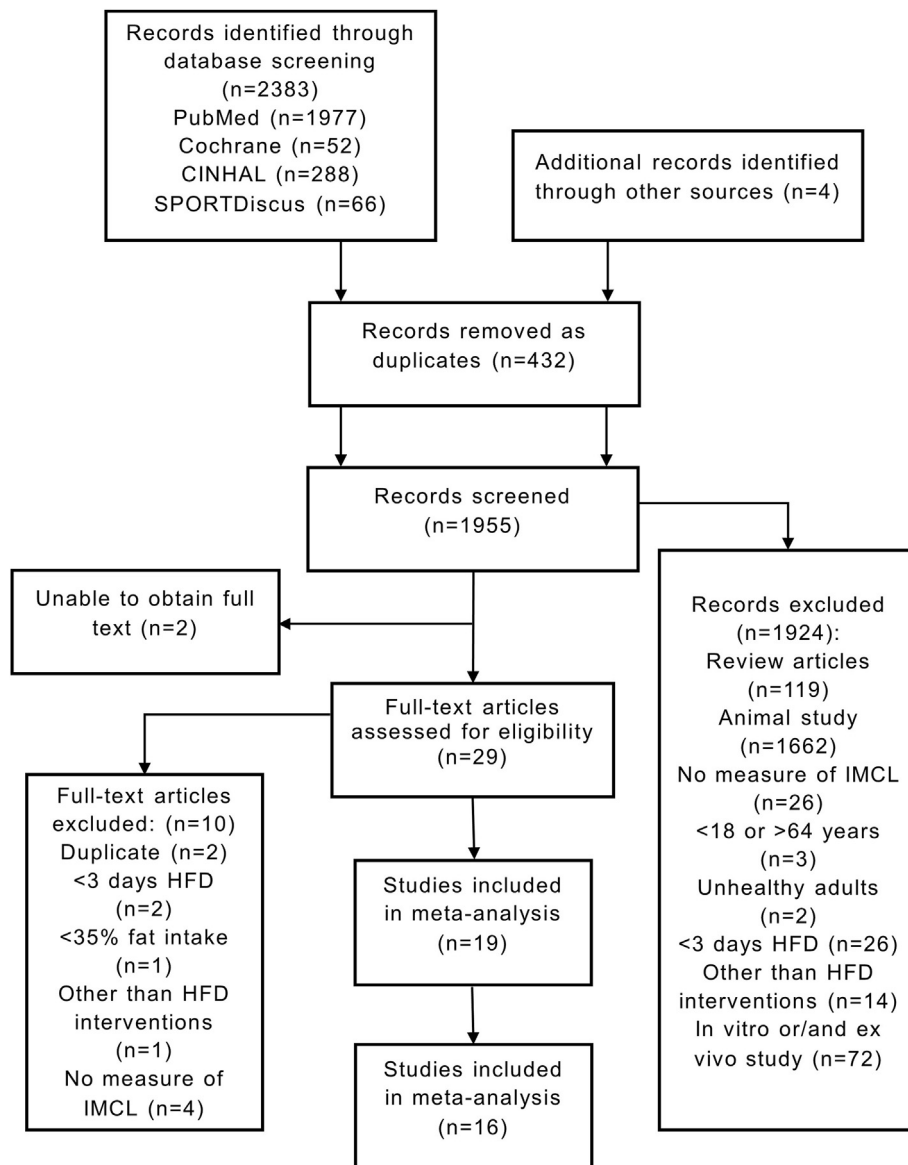


FIGURE 1. Flow chart of study selection. HFD, high-fat diet; IMCL, intramyocellular lipid.

small study effects ($\beta = 4.07$; 95% CI: 1.91, 6.22; $P < 0.001$; Figure 3), both of which may indicate a potential publication bias or small-study effect. Following HFD, there was an increase in IMCL content in studies using the microscopy technique ($37.9\% \pm 37\%$), $^1\text{H-MRS}$ ($29.1\% \pm 16.5\%$), or Biochemical extraction ($23.1\% \pm 36.6\%$). However, meta-regression revealed that the determination of IMCL responses to HFD was not affected by the IMCL measurement technique ($P > 0.05$) (Table 2).

Circulating triglyceride, and NEFA

Fasting circulating concentration of TAG (SMD = -0.30 ; 95% CI: $-0.68, 0.09$; $P = 0.12$ Figure 4A) nor NEFA (SMD = -0.26 ; 95% CI: $-0.81, 0.28$; $P = 0.31$; Figure 4B) changed in response to HFD. There was considerable heterogeneity between studies for both circulating TAG and NEFA concentrations ($I^2 = 83.24\%$; $P < 0.001$ and $I^2 = 91.26\%$; $P < 0.001$, respectively). The magnitude of heterogeneity was not affected by the study

characteristics as covariates in the meta-analysis. However, physical activity status contributed to changes in circulating TAG concentration ($P = 0.03$).

Insulin sensitivity

HFD did not affect fasting concentrations of circulating insulin (SMD = 0.22 ; 95% CI: $-0.09, 0.53$; $P = 0.15$; Figure 5A) and glucose (SMD = 0.11 ; 95% CI: $-0.18, 0.40$; $P = 0.42$; Figure 5B). There was a substantial and considerable degree of heterogeneity across studies that reported fasting circulating insulin ($I^2 = 74.37\%$; $P < 0.001$) and glucose concentration ($I^2 = 71.26\%$; $P < 0.001$), respectively.

Seven studies used the hyperinsulinemic-euglycemic clamp technique to measure insulin sensitivity [6,8,18,41,42,46,47], and 4 used the HOMA-IR [7,9,18,45]. Insulin sensitivity decreased in response to a HFD, as determined by the hyperinsulinemic-euglycemic clamp (SMD = -0.35 ; 95% CI: $-0.52, -0.17$; $P = 0.003$; Figure 6A), and HOMA-IR (SMD = 0.51 ;

TABLE 1

Summary of baseline data and effects of the high-fat diet on intramyocellular lipid content in the intervention group

Author	N (sex)	Participant characteristics (age; y)	HFD total fat content	Intervention diet	Comparator diet total fat content	Study design	Duration (day)	Measurement techniques	Major findings (IMCL content)
Adochioet al. [36]	M = 11 F = 10	Healthy lean (28)	50%, 156 g	Hypercaloric HFD (+40%)	30%, 67 g	Crossover (3 phases)	5	¹ H-MRS	↑
Cornford et al. [45]	M = 7 F = 2	Healthy, non-obese, physically inactive (24)	35%, 155 g	Hypercaloric HFD	NC	Single-arm trial	14	Biochemical extraction	↔
Covington et al. [8]	M = 29	Healthy, physically active (27)	44%	Hypercaloric HFD (140%)	NC	Single-arm trial	56	Microscopy, Oil red O stain	↔
Gemmink et al. [46]	M = 12	Healthy, lean Caucasian, and South Asian (22)	94%	Hypercaloric HFD	NC	Single-arm trial	5	Biochemical extraction	↑
Hoppeler et al. [38]	M = 7	Well-trained runners (37)	41%	Hypercaloric HFD	18%	Crossover (2 phases)	28–31	Transmission electron microscopy	↑
Johannsen et al. [18]	M = 29	Healthy (27)	44%, 207 g	Hypercaloric HFD (+40%)	NC	Single-arm trial	56	¹ H-MRS	↔
Kadowaki et al. [42]	M = 21	Healthy, non-obese (25)	48%, 320 g	Hypercaloric HFD (+45%)	NC	Single-arm trial	6	¹ H-MRS	↑
Kakehi et al. [41]	M = 50	Healthy, physically active (23)	60%	Normocaloric HFD	NC	Single-arm trial	3	¹ H-MRS	↑
Kien et al. [37]	M = 12 F = 12	Healthy, non-obese (29)	42%	Normocaloric high-palmitic acid diet and high-oleic acid diet	34%	Randomized parallel-arm trial (2 arms)	7	Biochemical extraction	↔
Kiens et al. [43]	M = 10	Healthy, nonobese, physically active (36)	54%	Normocaloric HFD	43%	Randomized parallel-arm trial (3 arms)	28	Biochemical extraction	↑
Lundsgaard et al. [47]	M = 9	Healthy, moderately trained (23)	78%, 496 g	Hypercaloric HFD, hypercaloric LFD, and Normocaloric NFD	24%, 87 g	Randomized crossover (3 phases)	3	Biochemical extraction	↑
Schrauwen-Hinderling et al. [39]	M = 7	Healthy (25)	60%, 218 g	Normocaloric NFD followed by Normocaloric HFD	NC	Single-arm trial	7	¹ H-MRS	↑
Skovbro et al. [44]	M = 21 (I = 10, CON = 11)	Healthy, untrained (24)	58%, 177 g	Normocaloric HFD and	30%, 76 g	Randomized parallel-arm trial (2 arms)	17.5	Biochemical extraction	↔

(continued on next page)

TABLE 1 (continued)

Author	N (sex)	Participant characteristics (age; y)	HFD total fat content	Intervention diet	Comparator diet total fat content	Study design	Duration (day)	Measurement techniques	Major findings (IMCL content)
St-Onge et al. [40]	M and F = 24	Healthy overweight (44)	37%	Normocaloric NFD Normocaloric HFD, Normocaloric HPUFA, and Normocaloric NFD	31%	Randomized crossover (3 phases)	25	¹ H-MRS	↑
Suzuki et al. [6]	M = 42	Healthy, non-obese, and physically active (23)	60%	Normocaloric HFD	NC	Single-arm trial	3	¹ H-MRS	↑
Tsintzas et al. [9]	M = 9	Healthy overweight/obese, physically inactive (44)	49%	Hypercaloric HFD (+25%)	NC	Single-arm trial	14	Microscopy, LD540 stain	↔
Van Proeyen et al. [49]	M = 7	Healthy, physically active (21)	50%, 123 g	Hypercaloric HFD (+30%)	34%, 113 g	Randomized parallel-arm trial (3 arms)	42	Microscopy, Oil red O stain	↑
Vogt et al. [48]	M = 11	Healthy, duathletes (32)	53%, 192 g	Normocaloric HFD (+12%) and Normocaloric LFD	17%, 53 g	Randomized crossover (2 phases)	35	Transmission electron microscopy	↑
Whytock et al. [7]	M = 11 F = 2	Healthy, lean, physically active (23)	64%, 325 g	Hypercaloric HFD (+47%)	NC	Single-arm trial	7	Microscopy, BODIPY stain	↑

¹H-MRS, H¹ magnetic resonance spectroscopy; BODIPY, boron dipyrromethene; CON, control; F, female; HFD, high-fat diet; HPUFA, high-polyunsaturated fat; I, intervention; IMCL, intramyocellular lipids; LD540, Lipophilic fluorescent dye; LDL, low density lipoprotein; LFD, low-fat diet; M, male; NC, no comparator; NFD, normal-fat diet; BODIPY; LD540.

↑: increased; ↔: unchanged. Because of missing data in some studies, values were not included.

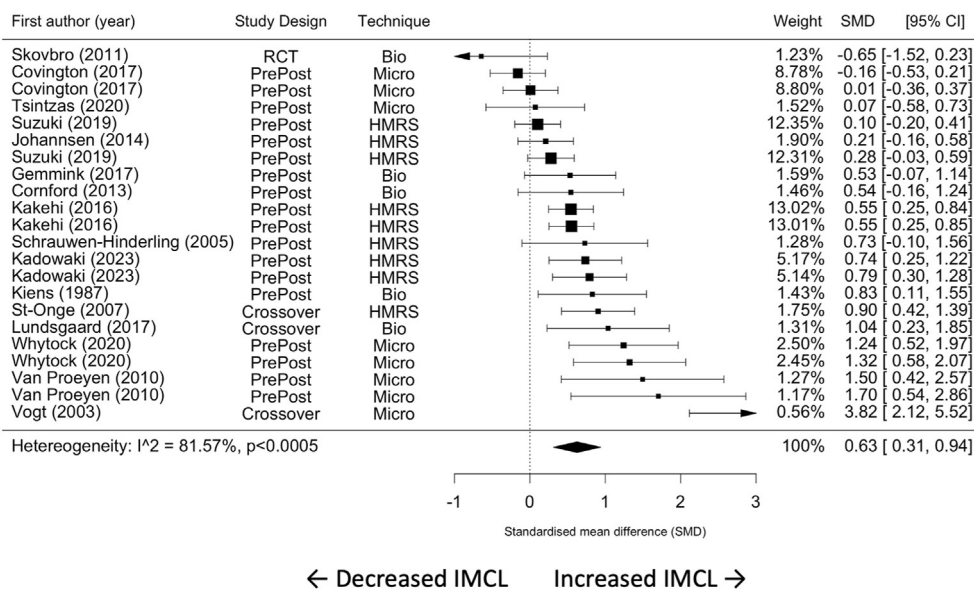


FIGURE 2. Forest plot of the effect of an HFD on IMCL content in healthy individuals. CI, confidence interval; HFD, high-fat diet; IMCL, intramyocellular lipid; SMD, standardized mean difference; ¹HMRS, H¹ magnetic resonance spectroscopy.

95% CI: 0.07, 0.95; *P* = 0.03; **Figure 6B**). The degree of heterogeneity might not be important between studies for hyperinsulinemic-euglycemic clamp (*I*² = 0.00%; *P* = 0.57) and HOMA-IR (*I*² = 0.00%; *P* = 0.64).

Meta-regressions

Meta-regressions for the effect of IMCL measurement techniques, study design, HFD duration, HFD total fat content, change in body mass, physical activity status, calorie intake, and change in IMCL content on outcomes are presented in **Table 2** and **Supplemental Table 2**. Physical inactivity and higher energy intake had a modifying effect on circulating insulin (*P* = 0.02; 0.01, respectively). Furthermore, higher energy intake was associated with a greater reduction in insulin sensitivity measured by HOMA-IR (*P* = 0.03). Additionally, physical

activity was associated with the change in circulating TAG concentration (*P* = 0.03). All other meta-regressions showed no significant modifying effects on outcomes.

Risk of bias

Six randomized studies were assessed by the RoB 2 tool, and 13 nonrandomized studies were assessed by the ROBINS-I tool. Of the 6 randomized studies, 5 studies were judged to raise some concerns about the overall risk of bias [37,40,44,47,49], and 1 was considered to have a high overall risk of bias because of not including a washout period [48]. Five randomized studies did not provide information on sources of recruitment [37,40,47–49]. Of the 13 nonrandomized studies, 2 studies were judged to have an overall low risk of bias [36,43], and 11 studies were considered to have a high overall risk of bias [6–9,18,38,39,41,

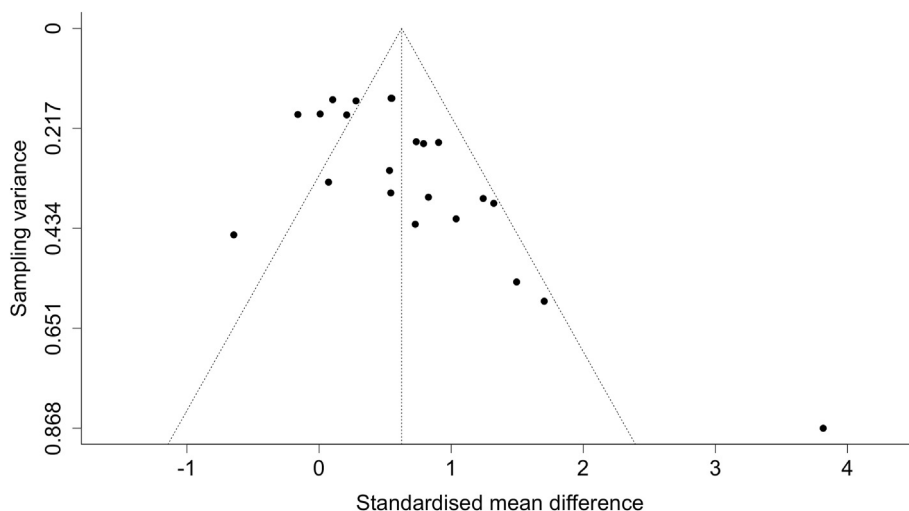


FIGURE 3. Funnel plot for the included studies in the meta-analysis.

TABLE 2
Summary of meta-regression results

Outcome	Covariate	Coefficient (95% CI)	P value	I ² (χ ² P value)
IMCL content	Measurement technique	0.41 (−0.33, 1.15)	0.26	83.92% (<0.0001)
	¹ H-MRS and Biochemical extraction ¹			
	Microscopy (n = 5)			
	Microscopy and Biochemical extraction ¹	−0.16 (−0.83, 0.51)	0.61	83.76% (<0.0001)
	¹ H-MRS technique (n = 6)			
	¹ H-MRS and microscopy ¹	−0.21 (−0.95, 0.53)	0.55	83.81% (<0.0001)
	Biochemical extraction technique (n = 5)			
	Study design (n = 16)	−0.33 (−1.16, 0.49)	0.41	83.12% (<0.0001)
	RCT and crossover trials ¹			
	Pre-post trials			
	HFD duration (n = 16)	−0.001 (−0.01, 0.016)	0.84	83.5% (<0.0001)
	Fat content (n = 10)	0.001 (−0.005, 0.006)	0.83	84.68% (0.0001)
	Body mass (n = 13)	−0.07 (−0.22, 0.07)	0.29	84.15% (0.0001)
	Physical activity status (n = 12)	0.42 (−0.23, 1.09)	0.19	83.44% (<0.0001)
Inactive ¹				
Active				
Energy intake (n = 16)	−0.02 (−0.70, 0.65)	0.94	83.82% (<0.0001)	
Normocaloric diet ¹				
Hypercaloric diet				
Hyperinsulinemic-euglycemic clamp	IMCL (n = 7)	−0.38 (−1.09, 0.34)	0.23	0% (0.71)
HOMA-IR	IMCL (n = 4)	0.43 (−1.03, 1.88)	0.33	0% (0.96)

HFD, high-fat diet; HOMA-IR, homeostatic model assessment for insulin resistance; IMCL, intramyocellular lipids; n, number of effect estimates; RCT, randomised controlled trial; ¹H-MRS, H¹ magnetic resonance spectroscopy; 95% CI, 95% confidence interval; χ², chi-squared test.

¹ Reference category in the model.

42,45,46] because of not controlling for confounding variables in the analysis. Detailed judgments for each domain in each included study are presented in Figure 7A, B, and C.

Discussion

The purpose of this systematic review and meta-analysis was to provide novel evidence on the effect of ≥3-d HFD on IMCL content and the association between changes in IMCL content and insulin sensitivity in healthy individuals. Evidence suggests that 1) IMCL is accumulated in healthy individuals following an HFD with a duration of between 3 and 56 d; 2) IMCL accumulation was not associated with HFD total fat content, duration, physical activity status, or measurement technique; 3) insulin sensitivity is reduced following HFD, but this was not related to IMCL accumulation.

The increase in IMCL content showed a moderate effect size in response to HFD. This agrees with the findings of a 7-study meta-analysis, where IMCL content increased following HFD [50]. This confirms that IMCL accumulation is a means by which excess dietary lipids can be accommodated [51]. Our findings are potentially explained by acute adjustments within the skeletal muscle to an HFD, which include a rapid upregulation of IMCL synthesis rates in response to elevated fatty acid availability [52] and chronic adaptations that favor enhanced fatty acid transport [53,54] and IMCL storage [7].

A novel finding of the present meta-analysis was that HFD total fat content was not associated with the increase in IMCL in response to HFD. This suggests that consuming a diet >35% of total energy intake elicits an increase in IMCL content, but greater fat intake may not further increase IMCL accumulation. Meta-regression analysis also revealed that the duration of HFD

was not associated with the increase in IMCL content. This means that a longer duration of HFD, beyond a minimum of 3 d, does not further increase IMCL accumulation. Together, these novel findings suggest that after as little as 3-d of a modest over-supply of lipids, IMCL stores may reach capacity in healthy individuals and that there must be an alternative fate for the continued over-supply of lipids other than storage as IMCL. However, meta-regression analysis may lack sensitivity because of the small number of studies that examined the effect of total fat content and duration on IMCL responses to HFD. In addition, bias within individual studies, visual inspection of the funnel plot, and Egger’s test of the intercept were suggestive of small-study effects, which may correspond to issues with publication and/or reporting bias, among other issues. To validate these findings, more experimental research is required to examine the time course of IMCL responses to HFD with varying fat content and/or duration.

The present study found no change in fasting circulating concentrations of TAG and NEFA following HFD. This indicates that an over-supply of lipids through high-fat feeding is countered by increased clearance of circulating TAG and NEFA. In support, an unchanged serum TAG concentration following a 4-wk HFD in healthy individuals coincided with increased skeletal muscle lipoprotein lipase activity and IMCL content [43]. The maintenance of circulating TAG and NEFA at baseline concentrations, despite continued HFD, occurred without further expansion of the IMCL pool. This may be explained by an increased rate of IMCL turnover [5,55,56], fat oxidation [57,58], and/or lipid storage within other tissues, such as hepatic [59], and subcutaneous, visceral and/or intermuscular adipose tissues [60].

To our knowledge, the present meta-analysis is the first to assess the modifying effect of study characteristics on IMCL

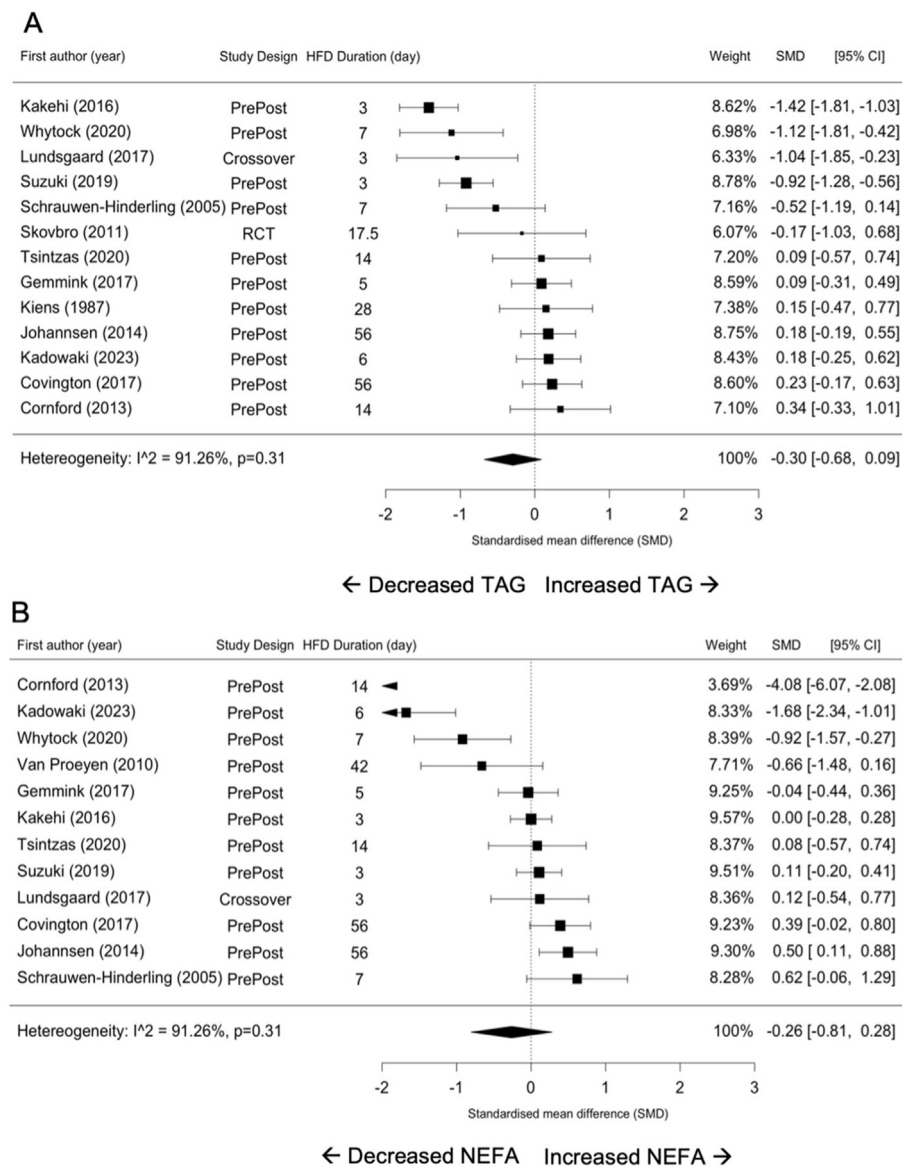


FIGURE 4. Forest plots of the effects of an HFD on circulating TAG (A) and NEFA (B). CI, confidence interval; HFD, high-fat diet; NEFA, nonesterified fatty acid; SMD, standardized mean difference; TAG, triacylglycerol.

responses to HFD. Endurance-trained individuals with a high IMCL content have a higher resting IMCL synthesis rate [61], IMCL turnover rate [62], and a greater oxidation rate of IMCL-derived fatty acids [63–65] compared with others. Therefore, we hypothesized that physically active individuals would have a greater capacity for lipid uptake, IMCL turnover, and fatty acid oxidation when challenged with HFD. This would enable lipid clearance despite a limited capacity for IMCL storage. Contrary to this hypothesis, meta-regression analysis showed no modifying effect of physical activity level on the IMCL response to HFD. However, the inclusion criteria employed by the present study may have resulted in a rather homogeneous sample in relation to physical activity. Further, physical activity was inconsistently measured and reported in some of the studies included, evidenced by the considerable between-study heterogeneity. As such, further research is required to elucidate the

effect of physical activity on the ability of skeletal muscle to tolerate and accommodate HFD.

Biochemical extraction, ¹H-MRS, and microscopy were the 3 measurement techniques used by studies to quantify IMCL content, and this may have contributed to the substantial heterogeneity we observed between studies in IMCL responses to HFD. For the first time using meta-regression analysis, we demonstrate no effect of the IMCL measurement technique on the assessment of change in IMCL content following HFD. This suggests that each of the 3 IMCL measurement techniques can be used to determine IMCL responses to HFD. This is surprising, as the IMCL measurement technique can affect the measurement of IMCL content at rest [66] and in the change in IMCL content following acute exercise [67]. Our finding may be explained by the significant increase in IMCL content following HFD being of a sufficient magnitude to minimize the effect of measurement

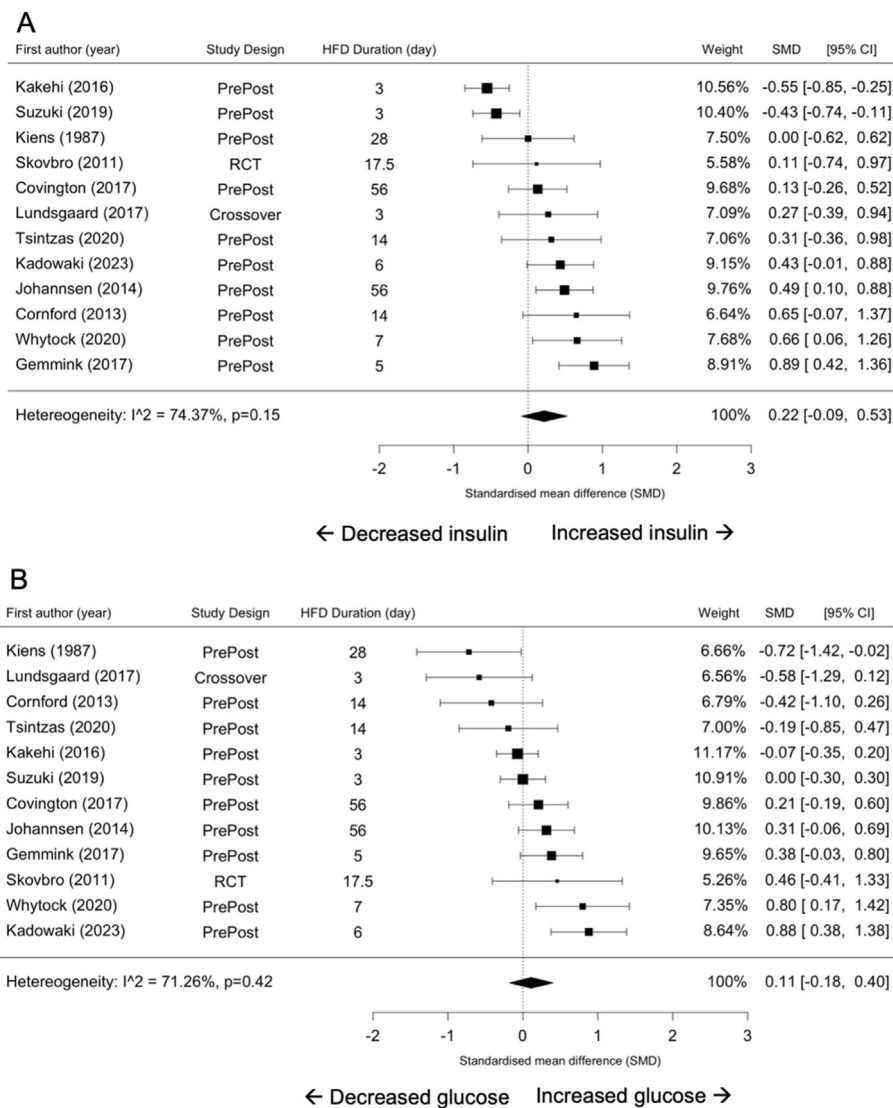


FIGURE 5. Forest plots of the effects of an HFD on circulating insulin (A) and glucose (B). CI, confidence interval; HFD, high-fat diet; SMD, standardized mean difference.

technique on assessing the change in IMCL content. However, caution is needed because of the relatively low number of included studies in this meta-regression.

We found a reduction in insulin sensitivity following HFD. However, meta-regression analysis revealed that the reduction in insulin sensitivity was not associated with the increase in IMCL content following HFD. This is unsurprising because it is well-accepted that a high-IMCL storage per se does not necessarily lead to insulin resistance [16]. A lower turnover of IMCL in physically inactive individuals [61] and those with excess adiposity [55] compared with more physically active individuals can result in the accumulation of fatty acid metabolites within skeletal muscle, such as diacylglycerol [68] and ceramide [69] and it is these which appear to exert the lipotoxic effect on skeletal muscle insulin signaling [70,71]. However, meta-regression analysis revealed that insulin sensitivity following HFD was not modified by physical activity status. In addition, of the included studies, evidence is mixed for an effect of HFD on diacylglycerol [7,9,47,49] and ceramide content [7–9,47,49] in healthy

individuals, suggesting further research is required. It is likely that excess lipid was taken up by subcutaneous, visceral, and/or intermuscular adipose tissues [60]. HFD-induced adipocyte hypertrophy has been shown to elicit tissue-specific [72,73] and whole-body [74] insulin resistance. In addition, intermuscular adipose tissue is negatively associated with insulin sensitivity [75] and may have contributed to the reduction in insulin sensitivity in the present study [76]. To what extent HFD affects intermuscular adipose tissue metabolism and insulin sensitivity in healthy individuals warrants further investigation.

This meta-analysis provides the most comprehensive and contemporary review to date, comprising 22 effect estimates from 16 studies assessing IMCL responses to HFD. It also included meta-regression analysis to investigate the potential sources of heterogeneity. Although this meta-analysis yields new, important evidence, some limitations to the current body of literature warrant consideration. Approximately 70% of the studies included were single-arm, pre-post studies, for which the risk of bias is inherently greater than the stronger study

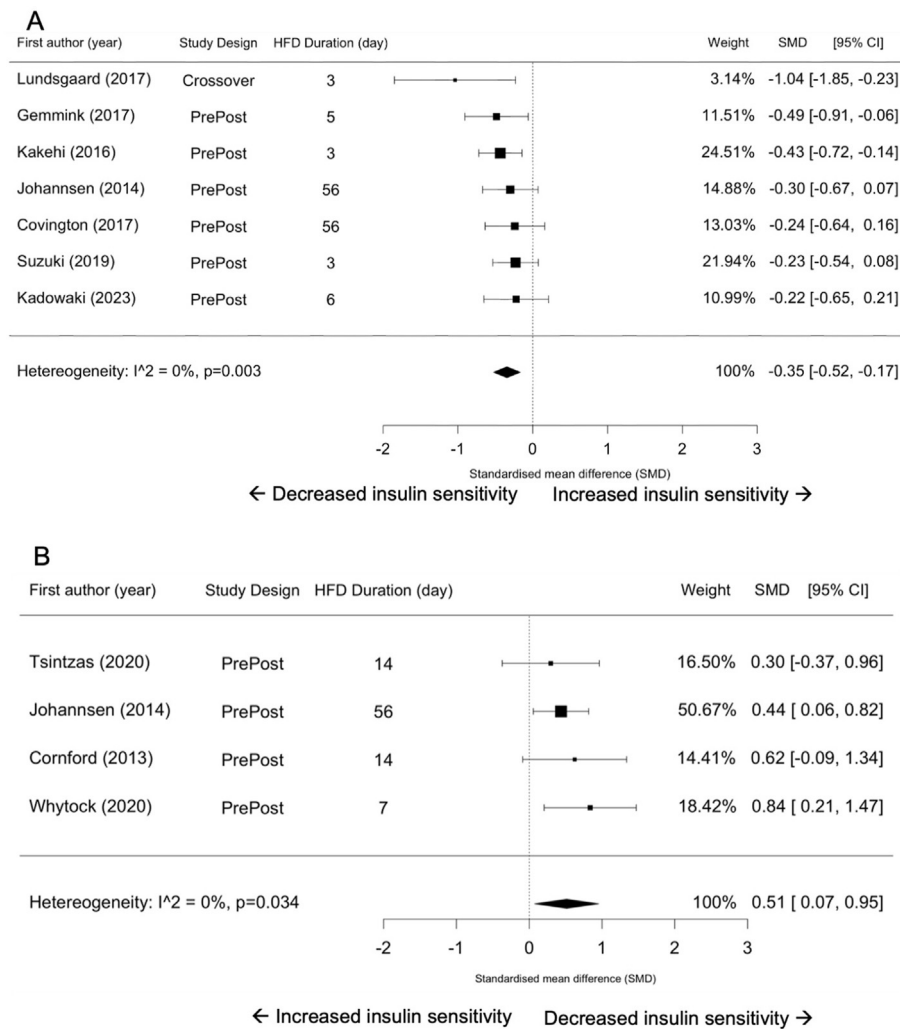


FIGURE 6. Forest plots of the effects of an HFD on hyperinsulinemic-euglycemic clamp (A) and HOMA-IR (B). CI, confidence interval; HFD, high-fat diet; SMD, standardized mean difference.

design of an RCT. Only ~6% of studies recruited female participants, and they did not report sex-specific effects of HFD on IMCL content. As it has been shown that females have higher IMCL accumulation in response to lipid infusion [77,78], it is possible that skeletal muscle lipid storage in response to HFD may differ between males and females. Because females are under-represented in this research, more studies are required to determine the sex-specific effects of an HFD on IMCL content. In RCT and crossover designs studies, the difference in fat content between the intervention and comparator diets was small. This could mean that the effect size of HFD on IMCL content was underestimated compared with normal-fat and low-fat diets. The effect of monounsaturated, polyunsaturated, and saturated fats on IMCL responses to HFD was not explored in the present meta-analysis. This is because only 1 study in the current meta-analysis investigated the effect of a high-polyunsaturated fat diet on IMCL content [40]. It is possible that IMCL content responds differently to a diet high in polyunsaturated, monounsaturated, or saturated fat [37,40]. The present study did not investigate the effect of ethnicity on the IMCL response to HFD

in different ethnic groups because of the lack of racial and ethnic diversity in the included studies. Future experimental studies should investigate the effect of HFD on IMCL content in different ethnic groups [79,80]. The present study explored IMCL responses to HFD in healthy adults aged <65 y, so our findings cannot be generalized to older adults and other individuals who possess insulin resistance, type 2 diabetes, and other chronic diseases.

In conclusion, this meta-analysis confirms that IMCL content is increased following an HFD in healthy individuals. Although an HFD does increase IMCL content, our data would suggest that the duration – beyond 3 d – and the fat content – beyond an intake exceeding 35% of daily energy from fat – do not influence IMCL accumulation. No effect of measurement techniques on the change in IMCL content was shown in response to HFD. Furthermore, a significant reduction in insulin sensitivity in response to HFD was observed, yet this reduction was not associated with the increase in IMCL content. Future well-designed trials are needed to improve the overall quality of evidence and the precision of the effect estimates.

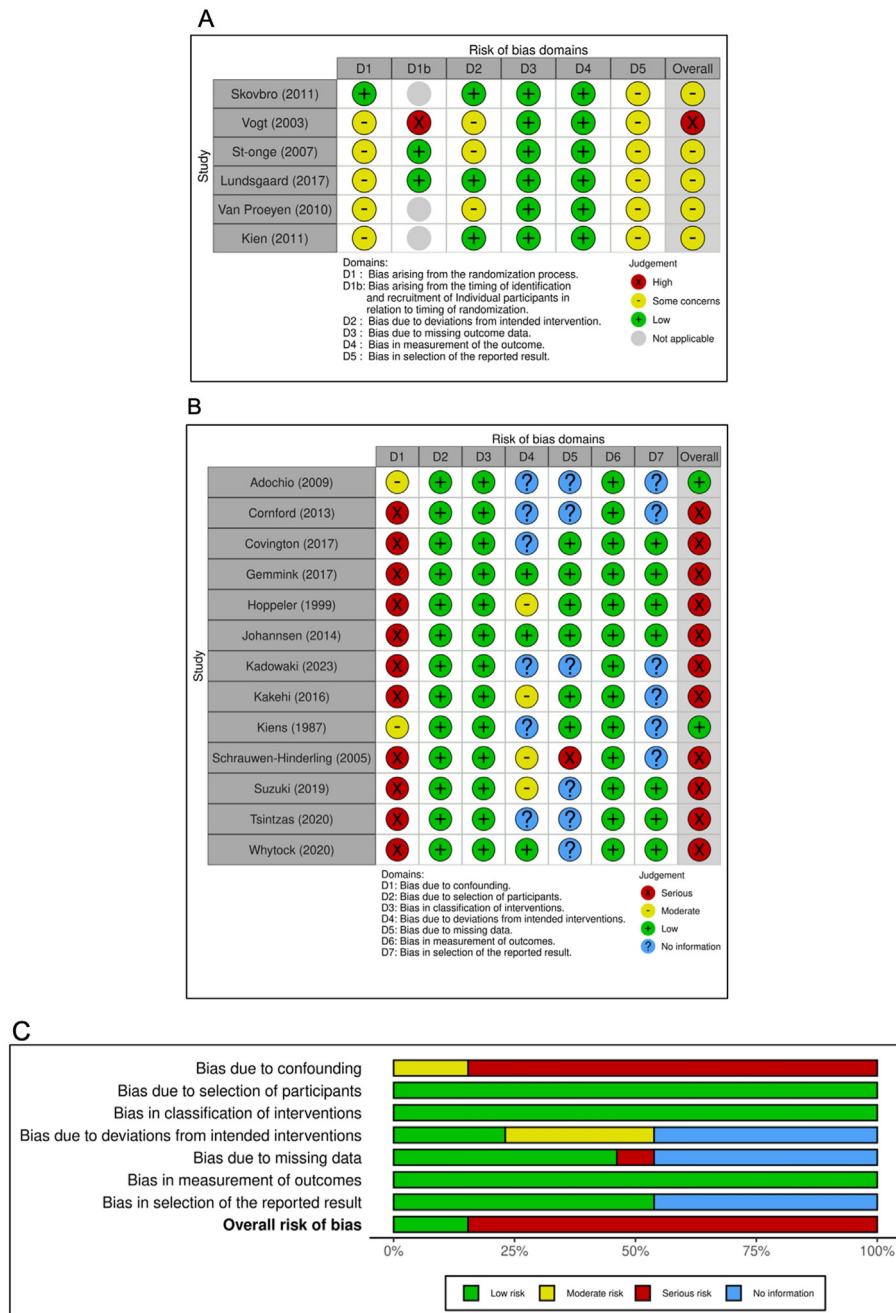


FIGURE 7. Assessment of risk of bias for randomized studies using the Cochrane RoB 2 tool (Traffic-light plot) (A), nonrandomized studies using the Cochrane ROBINS-I tool (Traffic-light plot) (B), and nonrandomized studies using the Cochrane ROBINS-I tool (Summary plot) (C). RoB, risk of bias.

Author contributions

The authors’ responsibilities were as follows – JA, OW, AS-K, AH, JM: were responsible for the conceptualization and development of the study; JA, KJ: performed the systematic searches, removed duplicates, screened the abstracts, assessed full texts for eligibility, and extracted outcome data; JA, AG: performed the risk of bias assessment; JA, STO: conducted the statistical analyses; JA, OW, AS-K, AH: interpreted the results; JA, OW: wrote the manuscript. OW: had primary responsibility for the final content, and all authors: read and approved the final manuscript.

Conflict of interest

The authors report no conflicts of interest.

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Data availability

Data will be made available upon request.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.tjn.2024.02.026>.

References

- [1] M.J. Arnold, M.C. Harding, A.T. Conley, Dietary guidelines for Americans 2020–2025: recommendations from the U.S. Departments of agriculture and health and human services, *Am. Fam. Physician*. 104 (5) (2021) 533–536.
- [2] V. Rakhra, S.L. Galappaththy, S. Bulchandani, P.K. Cabandugama, Obesity and the western diet: how we got here, *Mo, Med* 117 (6) (2020) 536–538.
- [3] Y. Kim, Y. Je, E.L. Giovannucci, Association between dietary fat intake and mortality from all-causes, cardiovascular disease, and cancer: a systematic review and meta-analysis of prospective cohort studies, *Clin. Nutr.* 40 (3) (2021) 1060–1070.
- [4] D.E. Kelley, B.H. Goodpaster, L. Storlien, Muscle triglyceride and insulin resistance, *Annu. Rev. Nutr.* 22 (2002) 325–346.
- [5] J.A. Kanaley, S. Shadid, M.T. Sheehan, Z. Guo, M.D. Jensen, Relationship between plasma free fatty acid, intramyocellular triglycerides and long-chain acylcarnitines in resting humans, *J Physiol* 587 (24) (2009) 5939–5950.
- [6] R. Suzuki, Y. Tamura, K. Takeno, S. Kakehi, T. Funayama, Y. Furukawa, et al., Three days of a eucaloric, low-carbohydrate/high-fat diet increases insulin clearance in healthy non-obese Japanese men, *Sci. Rep.* 9 (1) (2019) 3857.
- [7] K.L. Whytock, S.A. Parry, M.C. Turner, R.M. Woods, L.J. James, R.A. Ferguson, et al., A 7-day high-fat, high-calorie diet induces fibre-specific increases in intramuscular triglyceride and perilipin protein expression in human skeletal muscle, *J Physiol* 598 (6) (2020) 1151–1167.
- [8] J.D. Covington, D.L. Johannsen, P.M. Coen, D.H. Burk, D.N. Obanda, P.J. Ebenezer, et al., Intramyocellular lipid droplet size rather than total lipid content is related to insulin sensitivity after 8 weeks of overfeeding, *Obesity (Silver Spring)* 25 (12) (2017) 2079–2087.
- [9] K. Tsintzas, R. Jones, P. Pabla, J. Mallinson, D.A. Barrett, D.H. Kim, et al., Effect of acute and short-term dietary fat ingestion on postprandial skeletal muscle protein synthesis rates in middle-aged, overweight, and obese men, *Am. J Physiol. Endocrinol. Metab.* 318 (3) (2020) E417–E429.
- [10] H.E. Koh, J. Nielsen, B. Saltin, H.C. Holmberg, N. Ørtenblad, Pronounced limb and fibre type differences in subcellular lipid droplet content and distribution in elite skiers before and after exhaustive exercise, *J Physiol* 595 (17) (2017) 5781–5795.
- [11] J.A. Strauss, D.A. Shepherd, M. Macey, E.F.P. Jevons, S.O. Shepherd, Divergence exists in the subcellular distribution of intramuscular triglyceride in human skeletal muscle dependent on the choice of lipid dye, *Histochem. Cell Biol.* 154 (4) (2020) 369–382.
- [12] L.J. van Loon, Use of intramuscular triacylglycerol as a substrate source during exercise in humans, *J Appl. Physiol* 97 (4) (2004) 1170–1187, 1985.
- [13] V.B. Schrauwen-Hinderling, M.K. Hesselink, P. Schrauwen, M.E. Kooi, Intramyocellular lipid content in human skeletal muscle, *Obesity (Silver Spring)*. 14 (3) (2006) 357–367.
- [14] J.J. Deeks, J.P.T. Higgins, D.G. Altman, on behalf of the Cochrane Statistical Methods Group, Chapter 10: Analysing data and undertaking meta-analyses, in: J.P.T. Higgins, J. Thomas, J. Chandler, M. Cumpston, T. Li, M.J. Page, V.A. Welch (Eds.), *Cochrane Handbook for Systematic Reviews of Interventions* version 64, Cochrane, 2023 (updated August 2023).
- [15] D.A. Pan, S. Lillioja, A.D. Kriketos, M.R. Milner, L.A. Baur, C. Bogardus, et al., Skeletal muscle triglyceride levels are inversely related to insulin action, *Diabetes* 46 (6) (1997) 983–988.
- [16] B.H. Goodpaster, J. He, S. Watkins, D.E. Kelley, Skeletal muscle lipid content and insulin resistance: evidence for a paradox in endurance-trained athletes, *J Clin. Endocrinol. Metab.* 86 (12) (2001) 5755–5761.
- [17] C. Thamer, J. Machann, O. Bachmann, M. Haap, D. Dahl, B. Wietek, et al., Intramyocellular lipids: anthropometric determinants and relationships with maximal aerobic capacity and insulin sensitivity, *J Clin. Endocrinol. Metab.* 88 (4) (2003) 1785–1791.
- [18] D.L. Johannsen, Y. Tchoukalova, C.S. Tam, J.D. Covington, W. Xie, J.M. Schwarz, et al., Effect of 8 weeks of overfeeding on ectopic fat deposition and insulin sensitivity: testing the “adipose tissue expandability” hypothesis, *Diabetes Care* 37 (10) (2014) 2789–2797.
- [19] L. Shamseer, D. Moher, M. Clarke, D. Ghersi, A. Liberati, M. Petticrew, et al., Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation, *BMJ* 350 (2015) g7647.
- [20] L. Midão, A. Giardini, E. Menditto, P. Kardas, E. Costa, Polypharmacy prevalence among older adults based on the survey of health, ageing and retirement in Europe, *Arch. Gerontol. Geriatr.* 78 (2018) 213–220.
- [21] J.A. Berlin, Does blinding of readers affect the results of meta-analyses? University of Pennsylvania meta-analysis blinding study group, *Lancet* 350 (9072) (1997) 185–186.
- [22] Rohatgi, WebPlotDigitizer – Extract data from plots, images, and maps [internet], 2017. Available from: <https://automeris.io/WebPlotDigitizer/>. (Accessed 6 September 2023).
- [23] J.P.T. Higgins, J. Thomas, J. Chandler, M. Cumpston, T. Li, M. Page, et al., *Cochrane handbook for systematic reviews of interventions*, version 6.4 (updated August 2023) [Internet], Cochrane (2023). Available from: <http://www.training.cochrane.org/handbook>. (Accessed 12 November 2023).
- [24] J.A.C. Sterne, J. Savovic, M.J. Page, R.G. Elbers, N.S. Blencowe, I. Boutron, et al., RoB 2: a revised tool for assessing risk of bias in randomised trials, *BMJ* 366 (2019) 14898.
- [25] J.A. Sterne, M.A. Hernán, B.C. Reeves, J. Savović, N.D. Berkman, M. Viswanathan, et al., Robins-I: a tool for assessing risk of bias in non-randomised studies of interventions, *BMJ* 355 (2016) i4919.
- [26] J.P.T. Higgins, Cochrane Collaboration. *Cochrane handbook for systematic reviews of interventions*, 2nd ed., Wiley-Blackwell, Hoboken, NJ, 2020.
- [27] S.B. Morris, Estimating effect sizes from pretest-posttest-control group designs, *Organ. Res. Methods*. 11 (2) (2008) 364–386.
- [28] J.P.T. Higgins, T. Li, J. Deeks, Choosing effect measures and computing estimates of effect [internet], Cochrane (2021). Chapter 6. Available from: <http://www.training.cochrane.org/handbook>. (Accessed 8 October 2023).
- [29] R. Rosenthal, *Meta-analytic procedures for social research*, SAGE, Newbury Park, 1993.
- [30] S.T. Orange, K.M. Hicks, J.M. Saxton, Effectiveness of diet and physical activity interventions amongst adults attending colorectal and breast cancer screening: a systematic review and meta-analysis, *Cancer Causes Control* 32 (1) (2021) 13–26.
- [31] S.T. Orange, A.R. Jordan, J.M. Saxton, The serological responses to acute exercise in humans reduce cancer cell growth in vitro: A systematic review and meta-analysis, *Physiol. Rep.* 8 (22) (2020) e14635.
- [32] M. Borenstein, L.V. Hedges, J.P.T. Higgins, H.R. Rothstein, *Effect Sizes Based on Means* [Internet], 2009. Available from: <https://onlinelibrary.wiley.com/doi/abs/10.1002/9780470743386.ch4>. (Accessed 12 August 2023).
- [33] J. Cohen, *Statistical power analysis for the behavioral sciences*, 2nd ed., Routledge Academic, 1988.
- [34] J. Int’Hout, J.P. Ioannidis, G.F. Borm, The Hartung-Knapp-Sidik-Jonkman method for random effects meta-analysis is straightforward and considerably outperforms the standard DerSimonian-Laird method, *BMC Med. Res. Methodol.* 14 (2014) 25.
- [35] W. Van den Noortgate, J.A. López-López, F. Marín-Martínez, J. Sánchez-Meca, Three-level meta-analysis of dependent effect sizes, *Behav. Res. Methods*. 45 (2) (2013) 576–594.
- [36] R.L. Adochio, J.W. Leitner, K. Gray, B. Draznin, M.A. Cornier, Early responses of insulin signaling to high-carbohydrate and high-fat overfeeding, *Nutr. Metab. (Lond.)*. 6 (2009) 37.
- [37] C.L. Kien, K.I. Everingham, R.D. Stevens, N.K. Fukagawa, D.M. Muoio, Short-term effects of dietary fatty acids on muscle lipid composition and serum acylcarnitine profile in human subjects, *Obesity (Silver Spring)*. 19 (2) (2011) 305–311.
- [38] H. Hoppeler, R. Billeter, P.J. Horvath, J.J. Leddy, D.R. Pendergast, Muscle structure with low- and high-fat diets in well-trained male runners, *Int. J Sports Med.* 20 (8) (1999) 522–526.
- [39] V.B. Schrauwen-Hinderling, M.E. Kooi, M.K. Hesselink, E. Moonen-Kornips, G. Schaart, K.J. Mustard, et al., Intramyocellular lipid content and molecular adaptations in response to a 1-week high-fat diet, *Obes. Res.* 13 (12) (2005) 2088–2094.
- [40] M.P. St-Onge, B.R. Newcomer, S. Buchthal, I. Aban, D.B. Allison, A. Bosarge, et al., Intramyocellular lipid content is lower with a low-fat diet than with high-fat diets, but that may not be relevant for health, *Am. J Clin. Nutr.* 86 (5) (2007) 1316–1322.
- [41] S. Kakehi, Y. Tamura, K. Takeno, Y. Sakurai, M. Kawaguchi, T. Watanabe, et al., Increased intramyocellular lipid/impaird insulin sensitivity is associated with altered lipid metabolic genes in muscle of high responders to a high-fat diet, *Am. J Physiol. Endocrinol. Metab.* 310 (1) (2016) E32–E40.

- [42] S. Kadowaki, Y. Tamura, D. Sugimoto, H. Kaga, R. Suzuki, Y. Someya, et al., A short-term high-fat diet worsens insulin sensitivity with changes in metabolic parameters in non-obese Japanese men, *J Clin. Med.* 12 (12) (2023) 4084.
- [43] B. Kiens, B. Essen-Gustavsson, P. Gad, H. Lithell, Lipoprotein lipase activity and intramuscular triglyceride stores after long-term high-fat and high-carbohydrate diets in physically trained men, *Clin. Physiol.* 7 (1) (1987) 1–9.
- [44] M. Skovbro, R. Boushel, C.N. Hansen, J.W. Helge, F. Dela, High-fat feeding inhibits exercise-induced increase in mitochondrial respiratory flux in skeletal muscle, *J Appl. Physiol.* 110 (6) (2011) 1607–1614, 1985.
- [45] A.S. Cornford, A. Hinko, R.K. Nelson, A.L. Barkan, J.F. Horowitz, Rapid development of systemic insulin resistance with overeating is not accompanied by robust changes in skeletal muscle glucose and lipid metabolism, *Appl. Physiol. Nutr. Metab.* 38 (5) (2013) 512–519.
- [46] A. Gemmink, L.E. Bakker, B. Guigas, E. Kornips, G. Schaart, A.E. Meinders, et al., Lipid droplet dynamics and insulin sensitivity upon a 5-day high-fat diet in Caucasians and South Asians, *Sci. Rep.* 7 (2017) 42393.
- [47] A.M. Lundsgaard, K.A. Sjøberg, L.D. Høeg, J. Jeppesen, A.B. Jordy, A.K. Serup, et al., Opposite regulation of insulin sensitivity by dietary lipid versus carbohydrate excess, *Diabetes* 66 (10) (2017) 2583–2595.
- [48] M. Vogt, A. Puntschart, H. Howald, B. Mueller, C. Mannhart, L. Gfeller-Tuescher, et al., Effects of dietary fat on muscle substrates, metabolism, and performance in athletes, *Med. Sci. Sports Exerc.* 35 (6) (2003) 952–960.
- [49] K. Van Proeyen, K. Szulcfcik, H. Nielens, K. Pelgrim, L. Deldicque, M. Hesselink, et al., Training in the fasted state improves glucose tolerance during fat-rich diet, *J Physiol* 588 (21) (2010) 4289–4302.
- [50] S. Ahmed, D. Singh, S. Khattab, J. Babineau, D. Kumbhare, The effects of diet on the proportion of intramuscular fat in human muscle: A systematic review and meta-analysis, *Front. Nutr.* 5 (2018) 7.
- [51] C. Moro, J.E. Galgani, L. Luu, M. Pasarica, A. Mairal, S. Bajpeyi, et al., Influence of gender, obesity, and muscle lipase activity on intramyocellular lipids in sedentary individuals, *J Clin. Endocrinol Metab.* 94 (9) (2009) 3440–3447.
- [52] G. Boden, B. Lebed, M. Schatz, C. Homko, S. Lemieux, Effects of acute changes of plasma free fatty acids on intramyocellular fat content and insulin resistance in healthy subjects, *Diabetes* 50 (7) (2001) 1612–1617.
- [53] A. Bonen, S.S. Jain, L.A. Snook, X.X. Han, Y. Yoshida, K.H. Buddo, et al., Extremely rapid increase in fatty acid transport and intramyocellular lipid accumulation but markedly delayed insulin resistance after high fat feeding in rats, *Diabetologia* 58 (10) (2015) 2381–2391, 2015.
- [54] D. Cameron-Smith, L.M. Burke, D.J. Angus, R.J. Tunstall, G.R. Cox, A. Bonen, et al., A short-term, high-fat diet up-regulates lipid metabolism and gene expression in human skeletal muscle, *Am. J Clin. Nutr.* 77 (2) (2003) 313–318.
- [55] B.C. Bergman, L. Perreault, A. Strauss, S. Bacon, A. Kerege, K. Harrison, et al., Intramuscular triglyceride synthesis: importance in muscle lipid partitioning in humans, *Am. J Physiol. Endocrinol. Metab.* 314 (2) (2018) E152–E164.
- [56] M. Sacchetti, B. Saltin, T. Osada, G. van Hall, Intramuscular fatty acid metabolism in contracting and non-contracting human skeletal muscle, *J Physiol* 540 (1) (2002) 387–395.
- [57] P. Schrauwen, W.D. van Marken Lichtenbelt, W.H. Saris, K.R. Westerterp, Changes in fat oxidation in response to a high-fat diet, *Am. J Clin. Nutr.* 66 (2) (1997) 276–282.
- [58] P. Schrauwen, A.J. Wagenmakers, W.D. van Marken Lichtenbelt, W.H. Saris, K.R. Westerterp, Increase in fat oxidation on a high-fat diet is accompanied by an increase in triglyceride-derived fatty acid oxidation, *Diabetes* 49 (4) (2000) 640–646.
- [59] B. Ravikumar, P.E. Carey, J.E. Snaar, D.K. Deelchand, D.B. Cook, R.D. Neely, et al., Real-time assessment of postprandial fat storage in liver and skeletal muscle in health and type 2 diabetes, *Am. J Physiol. Endocrinol. Metab.* 288 (4) (2005) E789–E797.
- [60] L.M. Sparks, B.H. Goodpaster, B.C. Bergman, The metabolic significance of intermuscular adipose tissue: is IMAT a friend or a foe to metabolic health? *Diabetes* 70 (11) (2021) 2457–2467, 2021.
- [61] B.C. Bergman, L. Perreault, D.M. Hunerdosse, M.C. Koehler, A.M. Samek, R.H. Eckel, Increased intramuscular lipid synthesis and low saturation relate to insulin sensitivity in endurance-trained athletes, *J Appl. Physiol.* 108 (5) (2010) 1134–1141, 1985.
- [62] J. Lund, S.A. Helle, Y. Li, N.G. Løvsløtten, H.K. Stadheim, J. Jensen, et al., Higher lipid turnover and oxidation in cultured human myotubes from athletic versus sedentary young male subjects, *Sci. Rep.* 8 (1) (2018) 17549.
- [63] J. Calles-Escandón, M.I. Goran, M. O'Connell, K.S. Nair, E. Danforth Jr., Exercise increases fat oxidation at rest unrelated to changes in energy balance or lipolysis, *Am. J Physiol.* 270 (6 Pt. 1) (1996) E1009–E1014.
- [64] J.R. Berggren, K.E. Boyle, W.H. Chapman, J.A. Houmard, Skeletal muscle lipid oxidation and obesity: influence of weight loss and exercise, *Am. J Physiol. Endocrinol. Metab.* 294 (4) (2008) E726–E732.
- [65] B.H. Goodpaster, A. Katsiaras, D.E. Kelley, Enhanced fat oxidation through physical activity is associated with improvements in insulin sensitivity in obesity, *Diabetes* 52 (9) (2003) 2191–2197, 2003.
- [66] H. Howald, C. Boesch, R. Kreis, S. Matter, R. Billeter, B. Essen-Gustavsson, et al., Content of intramyocellular lipids derived by electron microscopy, biochemical assays, and ¹H-MR spectroscopy, *J Appl. Physiol.* 92 (6) (2002) 2264–2272, 1985.
- [67] J.R. Stokic, G. Abbott, K.F. Howlett, D.L. Hamilton, C.S. Shaw, Intramuscular lipid utilization during exercise: a systematic review, meta-analysis, and meta-regression, *J Appl. Physiol.* 134 (3) (2023) 581–592, 1985.
- [68] B.C. Bergman, D.M. Hunerdosse, A. Kerege, M.C. Playdon, L. Perreault, Localisation and composition of skeletal muscle diacylglycerol predicts insulin resistance in humans, *Diabetologia* 55 (4) (2012) 1140–1150.
- [69] F. Amati, J.J. Dubé, E. Alvarez-Carnero, M.M. Edreira, P. Chomentowski, P.M. Coen, et al., Skeletal muscle triglycerides, diacylglycerols, and ceramides in insulin resistance: another paradox in endurance-trained athletes? *Diabetes* 60 (10) (2011) 2588–2597.
- [70] L. Perreault, S.A. Newsom, A. Strauss, A. Kerege, D.E. Kahn, K.A. Harrison, et al., Intracellular localization of diacylglycerols and sphingolipids influences insulin sensitivity and mitochondrial function in human skeletal muscle, *JCI Insight* 3 (3) (2018).
- [71] B.C. Bergman, B.H. Goodpaster, Exercise and muscle lipid content, composition, and localization: influence on muscle insulin sensitivity, *Diabetes* 69 (5) (2020) 848–858.
- [72] M. Lavau, S.K. Fried, C. Susini, P. Freychet, Mechanism of insulin resistance in adipocytes of rats fed a high-fat diet, *J Lipid. Res.* 20 (1) (1979) 8–16.
- [73] M.S. Wiedemann, S. Wueest, F. Item, E.J. Schoenle, D. Konrad, Adipose tissue inflammation contributes to short-term high-fat diet-induced hepatic insulin resistance, *Am. J Physiol. Endocrinol. Metab.* 305 (3) (2013) E388–E395.
- [74] Y.S. Lee, P. Li, J.Y. Huh, I.J. Hwang, M. Lu, J.I. Kim, et al., Inflammation is necessary for long-term but not short-term high-fat diet-induced insulin resistance, *Diabetes* 60 (10) (2011) 2474–2483.
- [75] B.H. Goodpaster, F.L. Thaete, D.E. Kelley, Thigh adipose tissue distribution is associated with insulin resistance in obesity and in type 2 diabetes mellitus, *Am. J Clin. Nutr.* 71 (4) (2000) 885–892, 2000.
- [76] S. Sachs, S. Zarini, D.E. Kahn, K.A. Harrison, L. Perreault, T. Phang, et al., Intermuscular adipose tissue directly modulates skeletal muscle insulin sensitivity in humans, *Am. J Physiol. Endocrinol. Metab.* 316 (5) (2019) E866–E879.
- [77] L. Høeg, C. Roepstorff, M. Thiele, E.A. Richter, J.F. Wojtaszewski, B. Kiens, Higher intramuscular triacylglycerol in women does not impair insulin sensitivity and proximal insulin signaling, *J Appl. Physiol.* 107 (3) (2009) 824–831, 1985.
- [78] M.C. Devries, Sex-based differences in endurance exercise muscle metabolism: impact on exercise and nutritional strategies to optimize health and performance in women, *Exp. Physiol.* 101 (2) (2016) 243–249.
- [79] S. Lee, C. Boesch, J.L. Kuk, S. Arslanian, Effects of an overnight intravenous lipid infusion on intramyocellular lipid content and insulin sensitivity in African-American versus Caucasian adolescents, *Metabolism* 62 (3) (2013) 417–423.
- [80] S. Lee, N. Guerra, S. Arslanian, Skeletal muscle lipid content and insulin sensitivity in Black versus White obese adolescents: is there a race differential? *J Clin. Endocrinol. Metab.* 95 (5) (2010) 2426–2432.