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Citation:

Beggs, CB and Knibbs, LD and Johnson, GR and Morawska, L (2015) Environmental contamination and hospital-acquired infection: factors that are easily overlooked. *Indoor air*, 25 (5). 462 - 474. ISSN 0905-6947 DOI: <https://doi.org/10.1111/ina.12170>

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## **Environmental contamination and hospital acquired infection: factors that are easily overlooked**

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## **Environmental contamination and hospital acquired infection: factors that are easily overlooked**

### **Abstract**

There is an ongoing debate about the reasons for, and factors contributing to healthcare-associated infection (HAI). Different solutions have been proposed over time to control the spread of HAI, with more focus on hand hygiene than on other aspects such as preventing the aerial dissemination of bacteria. Yet, it emerges that there is a need for a more pluralistic approach to infection control; one that reflects the complexity of the systems associated with HAI, and involves multidisciplinary teams including hospital doctors, infection control nurses, microbiologists, architects, and engineers with expertise in building design and facilities management. This paper reviews the knowledge base on the role that environmental contamination plays in the transmission of HAI, with the aim of raising awareness regarding infection control issues that are frequently overlooked. From the discussion presented in the paper it is clear that many unknowns persist regarding aerial dissemination of bacteria, and its control via cleaning and disinfection of the clinical environment. There is a paucity of good quality epidemiological data, making it difficult for healthcare authorities to develop evidence-based policies. Consequently, there is a strong need for carefully designed studies to determine the impact of environmental contamination on the spread of HAI.

## **Environmental contamination and hospital acquired infection: factors that are easily overlooked**

### **1.0 Introduction**

In recent years there has been awareness that microbial contamination of the clinical environment may contribute to the spread of healthcare-associated infection (HAI) (Dancer et al. , 2009, Dancer, 2004, Dancer, 2008, Boyce et al. , 1997). Consequently, there has been increased emphasis on surface disinfection and ward cleaning, with some authorities placing a statutory obligation on hospitals to ensure that the clinical environment is clean and well maintained (N.P.S.A, 2007). While this has made the cleaning and disinfection of hospital wards a higher priority, the role that environmental contamination plays in the transmission of HAI is poorly understood. Indeed, there is little firm epidemiological evidence to support the widely-held and intuitive belief that cleaner hospitals result in fewer infections (Dancer et al., 2009, Dancer, 2008, Rhame, 1998). There is however, considerable evidence that the touching of contaminated surfaces by healthcare workers (HCWs) frequently results in the transient colonization of hands or gloves (Duckro et al. , 2005, Boyce et al., 1997, Hayden et al. , 2008, Bhalla et al. , 2004, Ray et al. , 2002), suggesting that there is a link between surface contamination and transient colonisation of the hands of HCWs. In fact, numerous studies have implicated contamination of the clinical environment in outbreaks of Gram-positive (Malamou-Ladas et al. , 1983, Hota, 2004, Fawley and Wilcox, 2001, Kumari et al. , 1998, Hardy et al. , 2006, Cotterill and al., 1996) and Gram-negative (Beggs et al. , 2006a, Das et al. , 2002, Weernink et al. , 1995, Allen and Green, 1987, Sherertz and Sullivan, 1985, McDonald et al. , 1998, Breathnach et al. , 2012, Engelhart et al. , 2002) bacterial infection. However, while much effort has been expended investigating environmental contamination and HAI, most of this has been detective work undertaken in response to specific hospital outbreaks. By comparison, very few controlled trials have been undertaken, with the result that the contribution made by environmental contamination to the overall body of HAI is difficult to quantify and characterize. It is therefore difficult to address with confidence, even basic questions regarding the cleanliness of hospitals. For example, it is not known which ward surfaces should be cleaned or disinfected, and how often such cleaning is required in order to minimise HAI rates. Indeed, it is not known if heavily contaminated surfaces such as hospital ventilation ducts, (which can accumulate particulate debris to a depth of several millimetres) pose any threat to the safety of patients. Consequently,

healthcare authorities have great difficulty specifying meaningful performance criteria for general hospital cleanliness.

To explore the role of environmental contamination in the transmission of infection within healthcare facilities, this paper aims to highlight infection control issues relating to building design and facilities management that have been overlooked, but might be worthy of further investigation. We limit our scope to sources of bacterial and fungal infection, as these have been more extensively described in the literature than viral pathogens.

## **2.0 Gulf between disciplines**

Traditionally, infection control has been the sole preserve of hospital doctors, infection control practitioners and microbiologists. However in recent years other professionals such as engineers have become involved in infection control, primarily because of their expertise in building design, facilities management, and modelling airflows within and between room spaces. In addition, many commercial organizations have developed hygiene related products for use in the healthcare sector. This increased activity has led to new insights into the transmission of some HAIs. However, while engineers and physical scientists have been able to make significant contributions to the infection control knowledge base, a gulf in thinking still exists between these professionals and hospital clinicians. This gulf reflects the various approaches inherent in these different occupations and is exemplified by a general belief amongst clinicians that the battle against HAI can be won only through greater hand hygiene compliance. Conversely, engineers and manufacturers tend to believe that gadgets and technical fixes might offer the optimum solution. This has polarized the debate, with clinicians tending to be very sceptical of infection control strategies not primarily focused on hand hygiene compliance, despite the fact that several studies have shown that the efficacy of hand hygiene measures can be severely limited by other factors (Beggs et. al. , 2008b, Beggs et. al. , 2006b, Talon et. al. , 2009, Silvestri et. al. , 2005, Beggs et. al. , 2009, DiDiodato, 2013). A middle ground between the two viewpoints that takes a 'whole of system' approach to HAI control would be able to capitalise on the expertise of all involved. Unfortunately, this has been conspicuous in its absence to date.

### 3.0 Complexity of the systems associated with HAI

HAIs arise from complex systems influenced by many factors, not the least of which are the biological characteristics of the infectious agents involved. In particular, the logistics associated with the management of patients and staff appear to be critical (Beggs et. al., 2006b). This complexity has largely been ignored by many in the infection control community, with the result that there has been a tendency to rely on single-measure strategies rather than taking a more holistic approach. However, the evidence suggests that single-measure strategies are not sufficient and that a more multi-faceted approach is required. Take for example the contrasting experiences of the health services in the Netherlands and UK. The Netherlands was quick to introduce a search-and-destroy (S&D) policy to counteract the emergence of drug resistant bacteria such as methicillin-resistant *S. aureus* (MRSA). The S&D policy was introduced as soon as cases of methicillin resistance were reported, although no official protocol existed until 1989 (Dekker and Van Den Broek, 2010). As the name implies, the Dutch authorities employed a comprehensive strategy that sought to isolate, contain and destroy MRSA whenever an infection was suspected or diagnosed. As such, the main focus of the strategy was on the screening and isolation of patients considered to be at increased risk for the carriage of MRSA (van Rijen and Kluytmans, 2009). All suspected patients were isolated and only released if cultures proved negative. When cultures were positive, the bacteria were first eradicated before the patient was released. In addition, hospital employees that had unprotected contact with MRSA positive patients were screened and prohibited from returning to work until they were culture negative (Dekker and Van Den Broek, 2010). This S&D strategy proved to be highly effective, maintaining the level of bacteraemia caused by MRSA at very low levels ( $\leq 1\%$ ) compared with other European countries that in some cases reached levels of up to 50% (Tiemersma et. al. , 2005). Similar screening and isolation strategies were adopted by healthcare trusts in the UK, but these were relaxed or abandoned in 1995 because of a lack of suitable isolation rooms, and also because ward closures and the cohorting of staff and patients caused considerable clinical disruption (Farrington et. al. , 1998). Instead, the UK focused on hand hygiene measures alone to fight MRSA – a policy encapsulated in the *Clean your hands* campaign (Stone et. al. , 2012). Unlike the Dutch experience, this policy resulted in a steady year-on-year increase in deaths associated with MRSA in England and Wales from 1993 to 2006, peaking at >1600 in 2006 (Pearson, 2009). Likewise, deaths associated with *Clostridium difficile* rose steadily from <1000

in 1999 to >8000 in 2007 (Pearson, 2009). While there is some evidence that the *Cleanyourhands* campaign, initiated in 2004, was responsible for stopping the steady rise of HAIs (Stone et. al., 2012), it was not until a raft of additional infection control measures were introduced around 2007 that HAI rates began to fall. With the introduction of care bundles (i.e. simple infection control guidelines for placing catheters, invasive lines and ventilator tubes); widespread deep cleaning of wards; cohorting of staff and patients; and improved screening of patients; MRSA and *C. difficile* infection rates fell by >50% (H.P.A., 2012b, H.P.A., 2012a). Collectively, these measures highlight the need for a more pluralistic approach to infection control; one that reflects the complexity of the systems associated with HAI.

#### **4.0 Aerial dissemination**

One interesting difference between the Dutch and British approaches to the control of MRSA is that the former assumed that MRSA can be disseminated by the airborne route, whereas the latter did not, as this route of transmission is generally not considered to be of great importance. Consequently, in the Netherlands MRSA isolation rooms are required to have an antechamber and to be negatively pressurized (van Rijen and Kluytmans, 2009), whereas in the UK, patients colonized with MRSA tend to be *barrier nursed* through placement in ward side rooms and the implementation of additional precautions to prevent the spread of infection. The differences between the approaches taken in the UK and Netherlands, highlights the tension that exists regarding the airborne transmission of infection in hospitals (Beggs, 2003a) – something that clinicians in many countries believe is of negligible importance compared to the spread of infection via the handborne route (Rhame, 1998, Ayliffe et. al. , 1999). Yet there is a large body of evidence, which suggests that both Gram-negative and Gram-positive pathogens are frequently disseminated by the aerial route in the clinical environment. Contaminated clothing and bedding of colonized patients release *S. aureus* into the air when disturbed (Solberg, 1965, Shiomori et. al. , 2002, Noble and Davies, 1965). Bed-making in particular liberates large numbers of particles (Roberts et. al. , 2006), many of which carry staphylococci into the air and these are then deposited on surfaces within the environment (Shiomori et. al. , 2001, Shiomori et. al., 2002, Noble, 1962). For example Rutala et al (Rutala et. al. , 1983) investigated a MRSA outbreak in a burn unit and found that MRSA accounted for 16% of all bacterial isolates sampled from the air, and 31% of the isolates cultured from elevated surfaces. In another study Shiomori et al

(Shiomori et. al., 2002), sampling the environment around MRSA colonized and infected patients under normal conditions, found an average of 4.7 cfu/m<sup>3</sup> MRSA-carrying particles in the air near infected patients; however during bed making this figure increased to 116 cfu/m<sup>3</sup>, confirming that this activity results in considerable aerosolization of particles containing staphylococci. Similarly, it has been demonstrated that *Acinetobacter* spp. (Houang et. al. , 2001, Beggs et. al., 2006a, Allen and Green, 1987, Das et. al., 2002, Gerner-Smidt, 1987, Obbard and Fang, 2003, Thornton et. al. , 2004) and *C. difficile* (Roberts et. al. , 2008) can be readily disseminated into the clinical environment by the aerial route. The different levels of recognition and acknowledgement of such evidence supporting airborne dissemination in the Netherlands and UK may have been a key factor in determining their relative success.

#### **4.1 Surface contamination due to aerial dissemination**

The extent to which aerial dissemination of bacteria contributes to surface contamination in hospitals has received little attention. Of the few studies undertaken, most have been carried out under controlled conditions in aerosol chambers (King et. al. , 2013, Hathway et. al. , 2007, Wong et. al. , 2010), with only a handful of studies linking particle dissemination with clinical activities (Roberts et. al., 2006, Hathway et. al. , 2011, Ayliffe et. al., 1999, Greene et. al. , 1960). Consequently, while it is known that aerial dissemination can result in surface contamination, there is little quantitative data on deposition rates and their variation, with which to make clinical judgments. It is however possible to make a rough estimate of particle deposition rates based on published data and using reasonable assumptions. For example, consider a 10 × 8 × 2.7 m ward room containing four patients (ignoring the presence of visitors, doctors, nurses, etc.), which experiences a ventilation rate of six air changes per hour. Based on earlier work by Roberts and Marks (Roberts and Marks, 1980), Milstone (Milstone, 2004) estimated that humans shed between 2 × 10<sup>8</sup> and 10 × 10<sup>8</sup> skin squamae per day, which equates to a combined average liberation rate of between 9259 and 46296 squamae per second for the four patients on the ward. If the air in the room space is well mixed and the squamae evenly distributed, then the calculated steady-state mean concentration of skin squamae in the air would be in the range 25719 to 128600 squamae/m<sup>3</sup>. If it is then conservatively estimated that each squamae carries 10 bacteria (Lundholm estimated that squamae frequently



carry >100 bacteria (Lundholm, 1982)), we arrive at a theoretical steady-state figure of 257190 to 1286000 bacteria/m<sup>3</sup> of air, which is of a similar order of magnitude to that found by Toivola et al (Toivola et. al. , 2004). If only 10% of the bacteria carrying particles deposit at an average rate of 2 mm/s (the settling velocity of an 8 µm particle), then a conservative estimate of the deposition rate would be of the order 51 to 257 bacteria/m<sup>2</sup> per second. Of course, in reality, the particles would not be evenly distributed throughout the room space, or shed at a constant rate; rather, they would be liberated into the air periodically in great numbers during bed making and other activities. This calculation should therefore be treated with caution. Notwithstanding this, this crude calculation does serve to illustrate an important and much over-looked point; namely, that aerial dissemination must be responsible for widespread surface contamination within the clinical environment. Evidence supporting this supposition comes from a 22-month surveillance study in which air vents and high horizontal surfaces were found to be contaminated with *C. difficile*, suggesting the aerial dissemination of isolates (Fawley and Wilcox, 2001, Fawley et. al. , 2003). Furthermore, outbreak strains of MRSA are frequently recovered from elevated surfaces (Rutala et. al., 1983) that are unlikely to have been touched by healthcare personnel, indicating that staphylococci must be transported through the air. Indeed, Boyce et al (Boyce et. al., 1997) found that patients colonized/infected with MRSA frequently contaminated room surfaces, with environmental contamination occurring in the rooms of 73% of MRSA-infected patients.

#### **4.2 Gram negative bioaerosols**

Immunocompromised patients in intensive care, high dependency, burns and haematology/oncology settings are particularly vulnerable to infection, many of which are associated with Gram-negative bacteria. These bacteria are strongly hydrophilic and are frequently cultured from sinks and wash hand basins in hospital wards (de Abreu et. al. , 2014, Kotsanas et. al. , 2013, Trautmann et. al. , 2001). There is increasing evidence that these microorganisms can become readily aerosolized. For example, Doring et al. (Doring et. al. , 1991) found sinks and basins on a hospital ward to act as reservoirs for *Pseudomonas aeruginosa*, with the result that the opening of water taps (faucets) generated bioaerosols containing *P. aeruginosa*, which contaminated the hands of HCWs during hand washing. An outbreak in a nursery was traced to a new air-conditioning system, from which *Acinetobacter* spp. Isolates were cultured (McDonald et. al., 1998). It was surmised that the outbreak

was caused by the dissemination of a bioaerosol generated by the contaminated air-conditioning unit. Contaminated air-conditioning units have also been implicated in outbreaks of *P. aeruginosa* infection (Pinna et al. , 2009). Interestingly Ryan et al (Ryan et al. , 2011) showed that by installing germicidal ultraviolet irradiation in the air conditioning system on a neonatal intensive care unit (NICU), it was possible to greatly reduce the incidence of ventilator-associated pneumonia (VAP).

The problem of Gram-negative bioaerosols becomes acute when the aerosols generated are specifically associated with medical equipment. With respect to this, the humidifiers and nebulizers associated with patient ventilator systems are particularly vulnerable (Jadhav et al. , 2013). When contaminated, they can 'inject' bioaerosols directly into the respiratory system, causing ventilator-associated pneumonia. Contaminated ventilator humidifiers and nebulizers have been implicated in outbreaks of *Pseudomonas* (Phillips, 1967, Redding and McWalter, 1980) and *Acinetobacter* (Cefai et al. , 1990, Ebenezer et al. , 2011). Portable medication nebulizers have also been implicated in infections of *Legionella pneumophila* (Mastro et al. , 1991), *Burkholderia Cepacia* (Hutchinson et al. , 1996) and *Stenotrophomonas maltophilia* (Denton et al. , 2003). These items of equipment require frequent washing/cleaning, and in the case of humidifiers, filling with sterile water, and are therefore vulnerable to contamination. Sinks and wash hand basins can act as reservoirs for Gram-negative pathogens, and a number of studies have implicated contaminated water supplies with increased risk of ventilator/nebulizer related infection (Mastro et al., 1991, Lucero et al. , 2011, Jarvis et al. , 2014).

## **5.0 Ward cleanliness**

Given that aerial dissemination of bacteria must be widespread in hospitals, why then is more attention not paid to this phenomenon? The simple answer to this question is that the clinical relevance of aerial dissemination is not well understood and therefore it is not considered a major problem. Outside of a few countries, notably the Netherlands and some Scandinavian countries, aerial dissemination of bacteria appears to have been largely ignored. One reason for this indifference is that the whole subject of ward cleanliness has generally been viewed as being of secondary importance compared with hand hygiene compliance. While the general public might associate visibly dirty wards with the transmission of MRSA infection, rather surprisingly there is relatively little epidemiological evidence that the environment is

important in endemic HAI (Rhame, 1998, Dancer et. al., 2009, Dancer, 2008, Maki et. al. , 1982, Collins, 1988, McGowan, 1981). Indeed, in a 2007 paper (Boyce et. al., 1997), the eminent microbiologist JM Boyce felt compelled to start his paper with the words: “*For several decades, there has been considerable controversy over whether or not contaminated environmental surfaces contribute to transmission of healthcare-associated pathogens.*” Given that contaminated surfaces can readily contaminate the hands of HCWs (Boyce et. al., 1997, Hayden et. al., 2008, Bhalla et. al., 2004, Ray et. al., 2002, Duckro et. al., 2005), one might wonder why there is any controversy. However, while it is relatively easy to show that colonized and infected patients can readily contaminate the clinical environment, it is much more difficult to demonstrate causality in the reverse direction. Consequently, epidemiological evidence supporting the link between ward cleanliness and HAI has been hard to obtain, with the result that healthcare authorities, hard-pressed by financial constraints, have tended to reduce the numbers of cleaners employed and the hours worked (Dancer, 2008). Furthermore, because the evidence base is sparse, cleaners specifications often focus on the cleaning of the most visible and widely-accepted locations like floors and toilets, rather than cleaning near-patient hand-touch sites, such as bed rails, bedside lockers, and infusion pumps, which are more likely to be of clinical importance (Dancer, 2008). As a result, cleaning of these near-patient surfaces may all too easily be overlooked.

In 2007, partly due to political pressure, but also due recognition that existing infection control policies had failed, the Department of Health in the UK rolled out a comprehensive hospital deep cleaning programme (D.O.H., 2008). At approximately the same time they also introduced a new national specification for hospital cleanliness (N.P.S.A, 2007) and imposed a statutory obligation on healthcare trusts to provide and maintain a clean clinical environment (N.P.S.A, 2007) – a noticeable departure from previous policy. Interestingly, the introduction of this policy coincided with a marked reduction in reported MRSA bacteraemia cases, which in England and Wales fell from 4451 in 2007-08 to 1114 in 2011-12 (H.P.A., 2012b) – something that was matched by a similarly large reduction in *C. difficile* associated infections (H.P.A., 2012a). This raises an obvious question about the extent to which the change in policy contributed to the reduction in HAI rates. However, this question is not easy to answer, because along with improved cleanliness, the Department of Health also introduced a raft of other measures, including improved strategies for placing and monitoring catheters and invasive lines, together with a continued push to improve hand hygiene compliance (*Cleanyourhands* campaign). Indeed, Stone et

al (Stone et. al., 2012) attributed the reductions in MRSA and *C. difficile* infection rates almost entirely to the *Cleanyourhands* campaign, which was commenced in 2004 - despite the fact that *C. difficile* infection rates did not start to fall until 2007. Noticeably, no mention was made, or analysis undertaken, of the contribution of improved ward cleanliness to the reduction in infection rates. Consequently, while intuitively one might feel that environmental contamination must influence HAI rates, concrete epidemiological evidence to this effect remains elusive due to the difficulty in disentangling the role of other factors. Notwithstanding this, there is evidence that hardy pathogens, such as *S. aureus* can be widely disseminated throughout the clinical environment via the hands of HCWs. Oelberg *et al* (Oelberg et. al. , 2000) using a viral DNA marker to inoculate a single telephone in a neonatal intensive care unit (NICU), observed that inanimate surfaces throughout the NICU rapidly became contaminated, with the number of positive sites peaking after only 8 hours. Similarly, Duckro *et al* (Duckro et. al., 2005) found vancomycin-resistant enterococci (VRE) to be rapidly disseminated around the clinical environment via HCW-surface interactions. Furthermore, Wilson et al (Wilson et. al. , 2004) observed a strong correlation between the presence of MRSA-colonized or –infected patients and air samples yielding MRSA in an ICU, suggesting widespread aerial dissemination. Given that contact with contaminated surfaces can readily lead to transient colonization of the hands of HCWs (Boyce et. al., 1997), there is good reason to believe that hospital cleanliness is likely to have an impact on HAI rates.

## **6.0 Hospital ventilation and duct contamination**

Most modern hospital buildings utilize mechanical ventilation air conditioning systems in order to maintain a comfortable environment for patients and staff. These systems contain large stretches of ductwork in which particulate matter can deposit and accumulate. Consequently, ducts in hospitals can become highly contaminated (see Figure 1). In recent years concern has been expressed about the risks posed by contaminated mechanical ventilation ductwork in hospital buildings. Yet, relatively little research has been carried out into the health risks associated with contaminated ventilation ductwork, particularly in healthcare facilities, with the result that little epidemiological evidence exists.



Figure 1. Typical example of a highly contaminated mechanical ventilation duct. Image courtesy of Total Ventilation Hygiene Pty Ltd and licensed for use in the HB2012 presentations and associated media.

While both supply and extract ducts may become heavily contaminated, in hospital buildings it is important to distinguish between the two, because the nature of the contamination is likely to be very different in the two types of ductwork. In supply ducts, because the air comes from a mixture of outdoor and filtered return air, fungal species are likely to predominate, whereas in the return air ducts, which extract from the ward spaces, contamination is likely to be predominately bacterial in nature. Of course, if the air is recirculated, as is the case in some healthcare facilities, then the bacterial pathogens from the ward space, such as MRSA, are likely to contaminate the supply duct and this might pose a greater hazard. Dust from occupied sections of buildings is largely comprised of skin squamae, and can accumulate in return ducts, especially when the air velocity is low (Batterman and Burge, 1995). It is therefore important when considering the subject of ductwork contamination to also consider the type of ventilation system in use, as this may have a bearing on the risk. Clearly if the recirculation of room air is permitted, then there is a greater likelihood of bacterial pathogens, being widely distributed around a healthcare facility via the mechanical ventilation system. Indeed, a number of studies relating to the transmission of

tuberculosis have shown this to be the case (Nardell et. al. , 1991, Beggs, 2003b, Houk, 1980).

Guidelines regarding the recirculation of air in healthcare settings vary greatly (Beggs et. al. , 2008a). For example, the American Institute of Architects (AIA) guidelines permit recirculation of ward air (A.I.A., 2001), whereas those for the United Kingdom in HTM 03 strongly discourage the use of recirculation systems (D.O.H., 2007). Because recirculation of air is permitted, in the United States the air supplied to patients in general wards must be first pre-filtered (minimum efficiency reporting value [MERV] 7, 30% dust spot efficiency), and then filtered to a MERV 14 or 15 standard (90% to 95% dust spot efficiency) before delivery to the ward space (A.S.H.R.A.E., 2003). This standard of filtration ensures 85% to 95% collection efficiency for 0.3 to 1.0  $\mu\text{m}$  particles and >90% efficiency for >1.0  $\mu\text{m}$  particles. Given that skin squamae are generally 4 to 25  $\mu\text{m}$  in size, this level of filtration should ensure that the air supplied to the ward space is relatively clean, despite the fact that a large proportion of this air may be recirculated. By comparison in the United Kingdom, where ward mechanical ventilation systems tend to be full fresh air, HTM 03 simply specifies the use of EU4 filters (>90% synthetic dust weight arrestance) in the supply air ducts to general ward spaces. Such filters are capable of removing the larger, heavier particles found in outdoor air. For critical care settings EU7 filters (80-90% dust spot efficiency) are specified, reflecting the higher perceived risk to patient safety in these areas (D.O.H., 2007).

## **6.1 Ductwork contamination**

Given that heavily contaminated ductwork such as that shown in Figure 1 can be found in hospital buildings, one might naturally assume that it poses a significant health hazard. However, the reality is that there are very little data directly relating environmental contamination of this type to adverse health effects (Kuehn, 2003). One reason for this might be that mechanical ventilation systems effectively act as a sink removing microbial particles from ward air – in effect, they act like a giant filter. If microbial particles are deposited within a ductwork system, then by definition they are removed from the air stream that enters the ward space. So in effect, the ductwork traps larger airborne particles preventing them from being distributed around the clinical environment. While the retention of particles might be considered beneficial, there is also a potential downside. If microbial particles from the ductwork become re-

suspended in the air for any reason, then they will be readily dispersed into the ward spaces. While relatively little is known about the re-suspension of bacterial matter, the same cannot be said for fungal spores that are uniquely adapted for aerial dissemination. Unlike bacterial matter, which generally requires the intervention of some mechanical force to create an aerosol, fungal spores are naturally disseminated by the airborne route, and so can easily re-enter the air stream within ventilation ducts. Given that hospital air conditioning and ductwork systems can become heavily contaminated with nosocomial fungal pathogens, such as *Aspergillus* species (Lutz et al. , 2003, Curtis et al. , 2005, Lentino et al. , 1982), there is reason to believe that ductwork colonized with fungal species might pose an infection risk, especially to immunocompromised patients (Buttner et al. , 1999).

There is evidence implicating contaminated ventilation systems with fungal infections in immunocompromised patients. Walsh and Dixon (Walsh and Dixon, 1989) cited contaminated ventilation systems as a common source of invasive aspergillosis, while Lentino et al (Lentino et al., 1982) implicated contaminated window mounted air conditioning units in an outbreak of pulmonary aspergillosis. In another study, Lutz et al (Lutz et al., 2003) identified mold contamination in an operating theatre air-handling system as the source of *Aspergillus* infections amongst post-surgical patients. They found that insulation material in variable-air-volume (VAV) units had become wet and had subsequently become colonized with several *Aspergillus* species. Insulation and filter media appear particularly vulnerable to fungal degradation when wet or under conditions of high humidity. Simmons and Crow (Simmons and Crow, 1995) found substantial growth of *Aspergillus* species on cellulosic filters at relative humidities >70%, and Maus et al (Maus et al. , 2001) observed significant growth of *Aspergillus niger* on used filters at relative humidities >85%.

With regard to the aerial dissemination of *Aspergillus* conidia, the case study described by Lutz et al (Lutz et al., 2003) highlights the importance of using terminal filtration in locations where immunocompromised patients might be vulnerable to infection. In this case, Lutz et al identified the fact that VAV units were mounted downstream of final filters as an issue of concern. When the insulation material in the ductwork became damp and degraded there was no barrier to filter the spores, and they were readily disseminated into the operating theatre. Given that *Aspergillus* conidia have diameters in the region 2-4  $\mu\text{m}$  (Lutz et al., 2003), somewhat smaller than skin squamae, it may be necessary to install high-performance terminal filtration

if the dissemination of spores is to be prevented – something highlighted in a study by Oren et al (Oren et. al. , 2001) who reported on an outbreak of pulmonary aspergillosis associated with construction activity. They found that airborne concentrations of *Aspergillus* species rose to a mean value of 15 cfu/m<sup>3</sup> in wards near a construction site. However, the installation of high-efficiency particulate air (HEPA) filters in hematological ward reduced the mean count to 0.18 cfu/m<sup>3</sup> and eliminated invasive pulmonary aspergillosis completely.

## **6.2 Ductwork cleaning**

Concerns regarding potential infection risks posed by contaminated ducting have led to a rise in the number of contractors offering specialist ductwork cleaning services to healthcare authorities. However, the evidence base of the efficacy of these measures in hospitals is limited. A somewhat larger body of evidence exists in relation to duct cleaning in residential and non-industrial commercial buildings, and this was recently reviewed with an aim of answering the question: *is ventilation duct cleaning useful?* (Zuraimi, 2010). The review, which employed strict inclusion criteria to assure that only peer reviewed, well designed and relevant papers were considered, came to several important conclusions. It firstly confirmed the existence of evidence that ventilation ducts are often contaminated with dust and provide conditions for microbiological growth, and that this happens under normal operating conditions. However, no field studies have conclusively correlated concentration of indoor pollution with duct contamination, despite controlled experiments showing that there is a plausible basis for this happening. The review also examined the available duct cleaning methodologies and showed that some of them are very efficient. However, again, it was unable to find consistent evidence that there is an improvement in indoor air quality after cleaning of the ducts. In fact, some of studies concluded that the opposite is the case, and that post-cleaning indoor concentrations are higher than pre-cleaning.

There is a disparity between the lack of evidence that duct cleaning can improve occupants' health or symptoms, and some suggestive evidence from epidemiological studies highlighting the association of dirty ducts with higher risk of symptoms. In general, the review demonstrated that that there is a need for balance between duct cleanliness and negative effects related to the process of cleaning. Nevertheless, the study demonstrated that the body of the evidence on many of the aspects discussed



is still small and identified specific areas requiring further research. While there are large differences between residential and commercial buildings in their purpose, design and operation, it is expected that the general conclusion on duct cleaning in such buildings is also generally applicable to hospital buildings, and that the knowledge gaps on the impact of duct cleaning in hospital building are even greater. Consequently, because the evidence base is sparse, healthcare authorities find it difficult to develop objective standards.

Most parts of ventilation systems can support microbial growth (Batterman and Burge, 1995), but frequently damp sections of ventilation air handling equipment and ducting most effectively promote mold growth, particularly *Aspergillus fumigatus* and *Aspergillus flavus*. This is especially true in areas where the primary role of the air handler is cooling, leading to substantial water condensation (Horner, 2006). These species are also present in accumulated dust inside ducts. There is evidence to suggest that mechanical brushing is more efficient at removing such dust from metal ducts and compressed air cleaning is more efficient for plastic ducts, and that reductions of 75 to 94% in surface dust levels can be achieved under field conditions (Holopainen et. al. , 2003). Of course, it is critical that the potential for resuspension of colonized dust and its liberation into supply air be minimized. Most mechanical brushing systems also incorporate a vacuum collection device, but there is potential for fugitive dust to reach indoor areas (Zuraimi, 2010). Chemical disinfection treatments (biocides, ozone, etc.) may be required to deal with substantial fungal and bacterial growth. However, these can pose a potential health risk in their own right (Zuraimi, 2010). The health risk-benefit balance for duct cleaning is not clear, at least in non-healthcare indoor environments (Zuraimi, 2010). Given the specific nature of healthcare settings, the large number of potential infection sources within them, and the susceptible nature of their occupants, it is plausible that carefully performed and appropriate duct cleaning could reduce HAI risk. However, the evidence base on this topic is very limited, and there is a strong need for well-performed studies linking duct cleaning with health outcomes. In the meantime, it is prudent to prevent or limit microbial contamination in the first instance, through the use of ultraviolet germicidal irradiation (UVGI) in air handlers and ducting, for example (Horner, 2006).

## 7.0 Discussion and Conclusions

The discussion above highlights the considerable hole in the knowledge base regarding the role that environmental contamination plays in the transmission of HAI. Not only is there no agreement on the risks posed by specific issues such as ductwork contamination, there is little quantifiable evidence regarding the benefits, or otherwise, of cleaning and disinfecting hospital wards, despite a general consensus that it is probably a good thing to do. Because it is difficult to quantify, the impact of hospital cleanliness is easily ignored. For example, the substantial fall in HAIs in the UK since 2007 has been attributed by some (Stone et. al., 2012), almost entirely to the *Cleanyourhands* campaign, despite the introduction of care bundles and hospital deep cleaning at approximately that time. Although hand hygiene is a key infection control measure of great importance, there is growing evidence that a multi-faceted approach is necessary. Evidence from several mathematical simulation studies (Beggs et. al., 2006b, Beggs et. al., 2008b, Beggs et. al., 2009) suggests that poor hand hygiene compliance is only one factor in the spread of HAI, and that other factors must be at work. In particular, the benefits of hand hygiene compliance appear to be threshold limited (Beggs et. al., 2008b, Beggs et. al., 2009). Indeed, a recent large Canadian study involving 166 acute care hospitals (DiDiodato, 2013) found that despite significant improvements in reported rates of hand hygiene compliance amongst healthcare personnel, no consistent reduction in either *C. difficile* infections or MRSA bacteremia was achieved. This prompted the study's author to state: *"This study supports the emerging evidence that once a threshold level of hand hygiene compliance is achieved, there is very little if any benefit to attempting to achieve higher rates of hand hygiene compliance among healthcare providers."* The UK experience since 2007 would tend to support this opinion. The introduction of a multi-faceted approach involving the introduction of: patient screening; cohorting of patients and nurses; careful use of antibiotics; improved placement and management of intravenous lines and catheters; improved management of ventilated patients; ward deep cleaning; and greater emphasis on hospital cleanliness; as well as improved hand hygiene, has led to a dramatic fall in MRSA and *C. difficile* infection rates. However, while great improvements have been made it is difficult to say which particular measures have been the most effective. So the contribution of improved ward cleanliness to the overall reduction in HAIs is difficult to quantify. Consequently, more research is needed to understand and quantify the role that ward cleaning plays in preventing HAIs.

One advance in recent years has been the trend towards a more multi-disciplinary approach to infection control. In particular, the involvement of engineers in infection control has led to advances in the application of technologies such as UVGI (Beggs and Sleigh, 2002, Noakes et. al. , 2004b, Noakes et. al. , 2004a, Beggs et. al. , 2000, Beggs, 2002, Cairns et. al. , 2001, Beggs et. al. , 2006c), negative air ionization (Kerr et. al. , 2006, Fletcher et. al. , 2007, Fletcher et. al. , 2008), and hydrogen peroxide terminal disinfection (Otter et. al. , 2006, Otter et. al. , 2007, Otter et. al. , 2008, Otter and French, 2009). While these technologies have merit, engineers can make the mistake of thinking that HAIs can be eliminated using a quick-fix technological solution. Indeed, many devices have not delivered reductions in HAI, primarily because their inventors failed to understand the complexity of the epidemiological systems associated with HAI. Having said this, if used appropriately as part of a multi-faceted approach to infection control, some of these environmental technologies may prove to be an important part of the solution. It is critical for engineers and others involved in technological solutions to bear in mind that the ultimate success or failure of an intervention is likely to depend more on the human element than the capability of the technology.

The simple calculation presented in section 4.1 above, suggests that aerial dissemination of bacteria may be a much greater problem than has been hitherto recognized. If staphylococci are being deposited onto surfaces from air at a rate  $>50$  bacteria/m<sup>2</sup> per second, as the calculation indicates, it would suggest that aerial dissemination may be the principal mechanism by which contamination of the clinical environment occurs. Although the clinical relevance of aerial dissemination is not known, there is good reason for believing that it may be important. Ayliffe et al (Ayliffe et. al., 1999) reported that sterile gauze and forceps laid on a horizontal surface, became readily contaminated by bacteria through aerial dissemination after bed making and curtain shaking. Das et al. (Das et. al., 2002) implicated heavily contaminated bed curtains in an outbreak of *Acinetobacter baumannii*, which when moved promoted the airborne spread of *Acinetobacter* species. Similarly, Weernink et al. (Weernink et. al., 1995) implicated feather pillows in the aerial dispersal of *Acinetobacter* species. Boyce et al (Boyce et. al., 1997) found that 42% of personnel who had no direct contact with MRSA patients, but had touched contaminated surfaces within the ward space, contaminated their gloves with MRSA. Furthermore, Noble et al. (Noble, 1981) found that the size distribution of particles containing *S. aureus* was approximately 4–25  $\mu\text{m}$ , which is roughly the size of skin squamae and

well in excess of the size of single *S. aureus* cells (i.e. about 1  $\mu\text{m}$  diameter). Noble et al. therefore surmised that most of the airborne *S. aureus* organisms were carried on skin squamae. Given that humans liberate  $>2 \times 10^8$  skin squamae into the air every day (Milstone, 2004), Noble et al. concluded that in many people a closed loop exists in which contaminated skin squamae are released into the air; they become impacted on the nasal turbinates; *S. aureus* grows on the nasal mucosa; hands then touch the nose and *S. aureus* bacteria are transferred to the skin; they colonize the skin and are ultimately disseminated back into the air on skin squamae.

The role of environmental contamination in the spread of Gram-negative bacterial infection is one that is becoming increasingly recognized. The ease with which contaminated sanitary fittings can lead to both the contamination of HCWs and the generation of aerosols containing Gram-negative bacteria is aptly illustrated by Doring et al. (Doring et al., 1991). They found 81% of sinks in a children's hospital to be contaminated with *P. aeruginosa*, something that contributed to the colonization of the hands of 42.5% of the HCWs on duty. Doring et al's study highlights the threat that sanitary fittings can pose to patient safety if they become contaminated. In critical care settings where patients are immunocompromised, the presence of contaminated sanitary fittings can easily lead to outbreaks of Gram-negative bacterial infection. Investigating a cluster of *Burkholderia cepacia* complex colonization in ventilated pediatric patients, Lucero et al (Lucero et al., 2011) identified tap water from hospital sinks as the likely mode of transmission. While they could not explain the exact mechanisms involved, the emergence of new cases stopped only after staff ended the practice of using tap water for oral and tracheostomy care. The link between clinical outcome and environmental contamination is further highlighted in an interesting study by Ryan et al (Ryan et al., 2011), who used ultraviolet light to irradiate cooling coils in an air-conditioning system serving a NICU. They found the cooling coils and condensate drain to be heavily colonized with Gram-negative bacterial species; as were the environmental surfaces and sink traps in the NICU. However, after six months of ultraviolet irradiation, both the air conditioning units and the environmental surfaces were culture negative. Furthermore, they found that patient tracheal colonization greatly reduced with the introduction of ultraviolet irradiation, as did the incidence of VAP. From this they surmised that airborne Gram-negative pathogens were being widely disseminated by the air-conditioning system and contributing to both surface contamination and patient colonization.

Given the magnitude of the HAI problem and the complexities of the systems involved, it is perhaps worth analyzing the ways in which microbes colonize the built environment (Kelley and Gilbert, 2013). Recently, researchers have sought to understand, from first principles, the microbial ecosystem that exists within hospitals, the so-called 'hospital microbiome' (Smith et. al. , 2013, Arnold, 2014). In the same way that antibiotics disrupt the normal microflora of the human body, constant attempts to 'sterilize' the clinical environment may in fact be removing a benign microbiome, which is capable of controlling and out-competing pathogenic species, only to replacing it with a microbial ecosystem that is more harmful to patients (Arnold, 2014). The fact that a sterile ward environment can become colonized with MRSA within 24-hours of admitting patients (Hardy et. al. , 2007), illustrates just how much 'nature abhors a vacuum'. Bacterial communities within indoor environments have been found to harbour microbial species not commonly found outdoors (Tringe et. al. , 2008). Kembel et al (Kembel et. al. , 2012) found that several bacterial taxa, commonly found in the human microbiome (including members of the families Burkholderiaceae, Pseudomonadaceae and Staphylococcaceae), were abundant in indoor air, especially in mechanically ventilated rooms, but nearly absent from outdoor air. Given that these species are commonly associated with humans as commensals or pathogens, they concluded that humans are important dispersal vectors for bacteria that colonize the built environment. Kembel et al (Kembel et. al., 2012) also found that building attributes, such as the source of ventilation air, relative humidity and temperature, correlated with the composition of indoor airborne bacterial communities, with the highest phylogenetic diversity found in the outdoor air and the lowest in rooms that were mechanically ventilated. This suggests that buildings can select for certain bacterial species, with the result that the indoor microbial ecosystem is less diverse and strongly influenced by the microflora of humans who spend as much as 90% of their lives indoors (Kelley and Gilbert, 2013). As such, it may be worth reappraising the way hospital buildings are designed. Reducing direct contact with the outdoor environment may not always be the best strategy for the management of bacterial pathogens (Kembel et. al., 2012). By creating an indoor environment that reflects the make-up of the outdoor air, it may be possible to create a more benign hospital microbiome. This challenges the assumption, held in many parts of the world, that hospitals should be sealed air-conditioned buildings, in which humidity and temperature are tightly controlled. It also presents a challenge to those seeking to minimize ventilation rates in order to save energy. However, it offers the possibility that if appropriate ventilation strategies can be identified, that move the hospital microbiome closer to that found in the outdoor

environment, then it may be possible to create an ecosystem which reduces the risk of patients acquiring a HAI.

Given increased emphasis on hospital hygiene in recent years, it is surprising that the whole issue of ventilation and its influence on the hospital microbiome has been largely overlooked. There is mounting evidence that the aerial dissemination of bacteria is a widespread phenomenon within the clinical environment. Yet little is known about how this influences the microbiome as a whole, or indeed the spread of HAI. Although environmental contamination has been implicated in some outbreaks of Gram-negative bacterial infection (Pinna et. al., 2009, McDonald et. al., 1998), the full extent to which it contributes to HAI is not known. For example, one might intuitively think that heavy contamination of air ductwork systems would pose a threat to patient safety. However, because few epidemiological studies have been undertaken specifically to investigate this subject, there is little evidence to substantiate this claim. Consequently, it is difficult to make any evidence-based decisions regarding optimum strategies to control HAI. Clearly, a better understanding of the microbial ecosystem within hospitals would be advantageous. A deeper understanding of the ways in which microbes disperse and colonize the clinical environment, together with the factors that influence this process, would provide a strong evidence base, which would be helpful in formulating future infection control strategies.

### **Acknowledgement**

At the Healthy Buildings 2012 conference in Brisbane in July 2012 a debate was conducted and attended by many experts on infection control and building design. This debate explored the role of environmental contamination in the transmission of infection within healthcare facilities. This paper arises from that debate and we are thankful to all those who contributed to it. In particular we thank the panel members, Tricia Coward, Yuguo Li, Jeremy Stamkos and Erica Stewart for their helpful contributions. This work was also supported by a Queensland University of Technology, IHBI Collaborative Research Development Grant titled, *How transmissible is influenza by the airborne route?* Luke Knibbs acknowledges an NHMRC Early Career (Australian Public Health) Fellowship (APP1036620).

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