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Fuel Use during Exercise at Altitude in Women with Glucose–Fructose Ingestion

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Short Title: Substrate oxidation in women at high altitude

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Abstract

Purpose: This study compared the co-ingestion of glucose and fructose on exogenous and endogenous substrate oxidation during prolonged exercise at terrestrial high altitude (HA) versus sea level, in women. **Method:** Five women completed two bouts of cycling at the same relative workload (55% W_{\max}) for 120 minutes on acute exposure to HA (3375m) and at sea level (~113m). In each trial, participants ingested 1.2 g.min⁻¹ of glucose (enriched with ¹³C glucose) and 0.6 g.min⁻¹ of fructose (enriched with ¹³C fructose) before and every 15 minutes during exercise. Indirect calorimetry and isotope ratio mass spectrometry were used to calculate fat oxidation, total and exogenous carbohydrate oxidation, plasma glucose oxidation and endogenous glucose oxidation derived from liver and muscle glycogen. **Results:** The rates and absolute contribution of exogenous carbohydrate oxidation was significantly lower at HA compared with sea level (ES>0.99, P<0.024), with the relative exogenous carbohydrate contribution approaching significance (32.6±6.1 vs. 36.0±6.1%, ES=0.56, P=0.059) during the second hour of exercise. In comparison, no significant differences were observed between HA and sea level for the relative and absolute contributions of liver glucose (3.2±1.2 vs. 3.1±0.8%, ES=0.09, P=0.635 and 5.1±1.8 vs. 5.4±1.7 grams, ES=0.19, P=0.217), and muscle glycogen (14.4±12.2% vs. 15.8±9.3%, ES=0.11, P=0.934 and 23.1±19.0 vs. 28.7±17.8 grams, ES=0.30, P=0.367). Furthermore, there was no significant difference in total fat oxidation between HA and sea level (66.3±21.4 vs. 59.6±7.7 grams, ES=0.32, P=0.557). **Conclusion:** In women, acute exposure to HA reduces the reliance on exogenous carbohydrate oxidation during cycling at the same relative exercise intensity.

Keys Words: Acute Hypoxia, Carbon Isotope, Exogenous Carbohydrate Oxidation, Liver Glycogen, Muscle Glycogen, Plasma Glucose Oxidation

1 **Introduction**

2
3 Endogenous stores of carbohydrate and fat are utilized as substrates to meet the metabolic demands
4 of the working muscle, as well as exogenous sources of carbohydrate if provided. The contribution
5 of carbohydrate and fat oxidation to energy expenditure at sea level is primarily related to the
6 intensity and duration of exercise, as well as the type and amount of carbohydrate ingested.
7 However, exposure to high altitude (HA) and the reduction in arterial oxygen saturation, is likely
8 to alter substrate oxidation during exercise compared with sea level. The complete oxidation of
9 carbohydrate requires less oxygen per mole of ATP synthesized compared with the oxidation of
10 free fatty acids (1) potentially leading to a greater reliance on carbohydrate oxidation in hypoxia.
11 However, the literature is inconclusive when comparing substrate oxidation during exercise
12 between hypoxia and normoxia (2). This heterogeneity may be explained by variations in study
13 design, such as participants' sex, method of exercise intensity determination (absolute vs. relative)
14 and whether carbohydrate was ingested or not (2).
15
16 Greater dependency on plasma glucose has been shown in men when comparing acute hypoxia
17 with normoxia, using the same absolute exercise intensity (3, 4). Conversely, when the exercise
18 intensity is normalized to the same relative exercise intensity the literature for men is equivocal
19 (2). These data either show an increase (5, 6) or no change in the respiratory exchange ratio (RER)
20 (7, 8). When an exogenous source of carbohydrate is provided on acute exposure to HA, men have
21 shown an increased reliance on endogenous carbohydrate oxidation (9), as well as a reduced
22 reliance on muscle glycogen and increased fat oxidation (10) during exercise at the same relative
23 intensity. However, it is known that sex influences substrate oxidation during exercise at sea level,
24 with women generally relying more on fat than carbohydrate as a fuel source compared with men

25 (11, 12). This has been attributed to women having a higher percentage of body fat (12), a greater
26 lipid content in muscles fibers (11) and better mobilization of fatty acids from subcutaneous
27 adipose tissue linked to cyclic changes in estrogen and progesterone (13). However, following the
28 provision of an exogenous source of glucose during exercise at sea level, most studies show no
29 statistical sex-differences in exogenous carbohydrate oxidation during exercise (14, 15) with one
30 exception (16). In the study by Riddell et al. (16), higher exogenous carbohydrate oxidation was
31 accompanied by a greater reduction in endogenous carbohydrate oxidation during exercise in
32 women compared with men. This is in contrast to other studies, which showed no difference (17)
33 or smaller reductions in endogenous or carbohydrate oxidation compared with men (14). Despite
34 this, there have only been limited studies assessing whole body carbohydrate or fat oxidation in
35 women at altitude.

36
37 In contrast to men, women have a lower RER, with increased free fatty acid availability when
38 exercising at a moderate relative exercise intensity (70% of altitude-related maximal oxygen
39 uptake ($\dot{V}O_{2\max}$)) on acute exposure to hypoxia (4,300m) compared with normoxia (18). These
40 women also showed no differences in substrate oxidation at HA between the early-follicular and
41 mid-luteal menstrual cycle phases when levels of estradiol and progesterone are high. Conversely,
42 a study, which recruited men and women, showed no change for either sex in RER during acute
43 hypoxic exposure (4100m) compared with normoxia when exercising at a matched relative
44 exercise intensity (50% of altitude-related $\dot{V}O_{2\max}$) (19). However, whole body carbohydrate
45 oxidation was increased at HA in both men and women following hypoxic exposure when using
46 the same absolute exercise intensity.

47 Considering the potential differences in fuel use between women and men at sea level, and what
48 has been documented for men at altitude, it is surprising that the fuel use responses to carbohydrate
49 ingestion during exercise at acute hypoxia have yet to be established in women. Thus, the purpose
50 of the present study was to compare the effects of co-ingesting ^{13}C glucose and ^{13}C fructose during
51 120 minutes of moderate intensity cycling exercise on exogenous and endogenous fuel use during
52 acute hypoxia (terrestrial high altitude) and normoxia (sea level) in women.

53

54 **Methods**

55 *Participants*

56 Seven women from the British military were recruited for this study, providing 86% power to
57 detect differences in the rate of exogenous carbohydrate oxidation, with an expected mean
58 difference of $0.2 \text{ g}\cdot\text{min}^{-1}$ between high altitude and sea level, assuming a standard deviation of 0.17
59 $\text{g}\cdot\text{min}^{-1}$ at an alpha of 0.05. However, only five completed the study due to attrition at points where
60 it was not possible to recruit additional participants. The five participants (age 25 ± 2 years, body
61 mass 61.7 ± 5.6 kg), were engaged in regular physical training (3-5 training days per week) and
62 considered to be physically fit but not elite athletes. Participants provided written informed consent
63 before the study, which was approved by the Ministry of Defence Research Ethics Committee
64 (Protocol 412/13). This cohort of participants were part of a larger research project and the design
65 of this study and the associated protocols have been described previously (10).

66

67 *Preliminary Testing*

68 Participants completed two maximal incremental cycle tests to volitional exhaustion to determine
69 their individual maximal workload ($W_{\text{max}}(20)$) and $\dot{V}\text{O}_{2\text{max}}$ on a bicycle affixed to a bicycle trainer
70 (Compu Trainer® Pro Lab, Racer Mate, USA), calibrated according to the manufacturer's

71 instructions. The first test was performed at sea level (altitude ~113m) with the second test
72 performed a week later during acute exposure to normobaric hypoxia (F_{iO_2} ~13.4% (considering
73 water vapour partial pressure (21) and daily fluctuations of barometric pressure) equivalent to
74 3375m (the reported altitude for the New Refuge Torino in the Italian Alps; P_{iO_2} 95.2 mmHg).
75 Oxygen uptake ($\dot{V}O_2$) and carbon dioxide production ($\dot{V}CO_2$) measurements were made
76 throughout using an online gas analysis system (Metalyser, Cortex, Germany), calibrated
77 following the manufacturer's instructions and were calculated using standard metabolic algorithms
78 (22) using the Haldane transformation. F_{iO_2} and F_{iCO_2} were measured continuously, rather than
79 assuming constants, thus correcting for changes in ambient conditions. This is important in a
80 normobaric chamber where the F_{iO_2} is reduced to simulate a given altitude. At sea level and
81 normobaric hypoxia participants achieved W_{max} values of 208.5 ± 34.4 W and 190.9 ± 31.3 W, and
82 $\dot{V}O_{2max}$ values of 44.7 ± 7.1 ml.kg⁻¹.min⁻¹ and 37.4 ± 5.9 ml.kg⁻¹.min⁻¹, respectively.

84 *Design of the Study*

85 Following the assessment of W_{max} , participants completed two experimental cycling trials for 120
86 minutes at 55% W_{max} , one at terrestrial HA (barometric pressure 506.7 ± 1.4 mmHg, P_{iO_2} : $96.3 \pm$
87 0.3 mmHg (New Refuge Torino, Alps, Italy)), as described previously (10) and another at sea level
88 seven weeks later. To control for menstrual cycle phases, the women were tested in either the
89 early-follicular or mid-luteal phase, which has no effect on fuel use at HA (18). Each cycling test
90 involved the ingestion of 1.8 g.min⁻¹ of carbohydrate (1.2 g.min⁻¹ of glucose (D-glucose, Thornton
91 and Ross Ltd, Huddersfield, UK) and 0.6 g.min⁻¹ of fructose (D-fructose, Danisco, Oy, Okta,
92 Finland) at regular intervals during exercise. Stock glucose (natural $\delta^{13}C$ abundance = -32.58 ‰)
93 and fructose (natural $\delta^{13}C$ abundance = -30.04 ‰), was enriched using 0.24 g of U-¹³C₆ D-glucose

94 (Cambridge Isotope Laboratories, Inc, Tewksbury, MA, USA), and 0.12g of U-¹³C₆ D-fructose
95 (Cambridge Isotope Laboratories, Inc), achieving a combined enrichment of $\delta^{13}\text{C} = +115.88 \%$.
96 All $\delta^{13}\text{C}$ measurements are quoted with reference to the internationally accepted standard for
97 carbon isotope measurements, Vienna Pee Dee Belemnite (VPDB). The ¹³C abundance of stock
98 glucose and fructose and ¹³C enrichment of spiked glucose and fructose was determined using
99 liquid chromatography coupled to isotope ratio mass spectrometry (LC-IRMS; Isoprime, Cheadle,
100 UK), using L-Fucose as an isotopic internal standard as previously described (23).

101

102 *Diet and physical activity before testing*

103 Participants recorded their food intake and activity patterns during the 72 hours before the first
104 experimental trial and were instructed to repeat the same diet and activity pattern before the
105 subsequent trial. Participants were required to refrain from any intense and/or prolonged physical
106 activity, alcohol or caffeine consumption in the 36 hours before each experimental trial. In
107 addition, they were asked to refrain from ingesting carbohydrates derived from plants which utilize
108 the C₄ photosynthetic cycle, in which there is higher natural abundance of ¹³C (e.g. maize derived
109 sugars) for the duration of the study. This precaution ensured that background ¹³CO₂ abundance
110 was less likely to be perturbed from oxidation of endogenous and dietary substrate stores from
111 naturally “enriched” C₄ origin. A standardized evening meal was consumed 12 hours before each
112 experimental trial (total 1443 kcal; 53% carbohydrate, 17% fat, 30% protein).

113

114 *Experimental Trials*

115 Each experimental trial was performed at 19 to 21°C following an overnight fast. Participants
116 repeated their trials at the same time of day, to avoid any influence of circadian variance. On arrival

117 a catheter (20 gauge Introcan Safety[®], B. Braun Medical Ltd, Sheffield, UK) was inserted into an
118 antecubital vein for regular blood sampling. After 20 minutes of acute exposure to each
119 environmental condition, peripheral oxygen saturation (SpO₂ (Nellcor N-20, Covidien, Dublin,
120 Ireland) was measured and resting blood samples were drawn for the analysis of plasma glucose,
121 serum insulin, serum free fatty acids, plasma lactate, plasma metanephrine and normetanephrine
122 concentrations, as well as plasma ¹³C glucose enrichment.

123
124 Participants then rested for 10 minutes whereby $\dot{V}O_2$ and $\dot{V}CO_2$ measurements were made using
125 an online gas analysis system (Metalyser, Cortex, Germany). For the measurement of ¹³C/¹²C in
126 expired CO₂, 12 ml samples of expired gas were collected in duplicate in Labco Exetainers[®]
127 (SerCon Ltd, Crewe, UK) via a mixing chamber (Jaeger, Germany).

128
129 After a 5 minute standardized warm up, which included the calibration of the bicycle trainer
130 (Compu Trainer Pro Lab, Racer Mate, USA), an initial bolus of the carbohydrate solution was
131 consumed (397ml). Participants then completed 120 minutes of cycling; 5 minutes at 40% W_{max},
132 5 minutes at 45% W_{max}, 5 minutes at 50% W_{max}, 105 minutes at 55% W_{max}. These workloads were
133 calculated from participants' sea level and normobaric hypoxic W_{max} for the sea level and HA
134 environments, respectively. Additional boluses (229ml) of the carbohydrate solution were
135 provided every 15 minutes throughout the 120 minute exercise period. Expired gas breath samples
136 were collected and measurements of $\dot{V}O_2$ and $\dot{V}CO_2$ were made every 15 minutes during exercise.
137 Samples of expired gas for ¹³CO₂ analysis were collected during the final 60 seconds of each
138 collection period. Samples for the analysis of plasma glucose, serum insulin, serum free fatty acids
139 and plasma lactate were drawn every 15 minutes, those for plasma ¹³C glucose enrichment were

140 drawn at 60, 90 and 120 minutes and those for plasma metanephrine and normetanephrine
141 concentrations were drawn at 60 and 120 minutes. Heart rate, rating of perceived exertion (RPE)
142 and SpO₂ was measured every 15 minutes during cycling exercise.

143

144 *Analyses*

145 Plasma and serum samples collected at HA were initially stored at -20°C until they were
146 transported back to the United Kingdom, where they were then stored at -80°C until analysis, as
147 per the samples collected at sea level. All samples were analysed in accordance with the procedures
148 described previously (10).

149

150 The ¹³C/¹²C ratio in expired CO₂ was determined using isotope ratio mass spectrometry (IRMS;
151 AP2003, GVI Instruments Ltd, Manchester, UK). The isotopic ratio ¹³C/¹²C is derived against
152 laboratory CO₂ (itself calibrated against VPDB) from the ion beam area ratio measurements with
153 correction of the small contribution of ¹²C¹⁶O¹⁷O at m/z 45; the Craig correction (24). The ¹³C/¹²C
154 ratio in plasma glucose was determined using LC-IRMS as described previously (23).

155

156 Oxidation rates of total fat, total carbohydrate, endogenous carbohydrate (liver and muscle),
157 plasma glucose and exogenous carbohydrate derived from glucose and fructose ingestion
158 combined, were calculated by indirect calorimetry ($\dot{V}O_2$ and $\dot{V}CO_2$) and stable isotope
159 measurements (¹³C/¹²C ratio in expired CO₂ and plasma glucose), as detailed below.

160

161

162

163 **Calculations**

164 Total CHO and fat oxidation ($\text{g}\cdot\text{min}^{-1}$) were computed from $\dot{V}O_2$ ($\text{L}\cdot\text{min}^{-1}$) and $\dot{V}CO_2$ ($\text{L}\cdot\text{min}^{-1}$)
 165 using stoichiometric equations (25), with the assumption that protein oxidation during exercise
 166 was negligible.

167
 168 The isotopic enrichment of the ingested glucose and fructose, (R_{exo}), was expressed in standard
 169 $\delta^{13}\text{C}$ units (‰) relative to VPDB (26). Exogenous carbohydrate oxidation derived from the
 170 combined ingestion of glucose and fructose (G_{exo} , grams) was computed by using equation 1 (27).

171
 172 Exogenous Carbohydrate Oxidation ($\text{g}\cdot\text{min}^{-1}$) = $\dot{V}CO_2 [(R_{\text{exp}} - R_{\text{ref}}) / (R_{\text{exo}} - R_{\text{ref}})] / k$ (1)

173
 174 where $\dot{V}CO_2$ is in litres per minute, R_{exp} is the measured isotopic composition in expired CO_2 , R_{ref}
 175 is the isotopic composition of expired CO_2 at rest before exercise and carbohydrate ingestion, R_{exo}
 176 is the measured isotopic composition of the exogenous glucose and fructose ingested, and k
 177 ($0.7426 \text{ l}\cdot\text{g}^{-1}$) is the rate adjusted value for the complete oxidation of glucose (28). The use of R_{ref}
 178 from expired CO_2 at rest is typical of studies in this area of research (9, 29) as the high ^{13}C -
 179 enrichment of exogenous glucose and fructose provides a strong signal in expired CO_2 . This
 180 cancels the confounding effects of relatively small fluctuations in background enrichment of
 181 expired CO_2 seen from rest to exercise following a Western European diet (30) on the calculation
 182 of exogenous carbohydrate oxidation. Endogenous carbohydrate oxidation was calculated by
 183 subtracting exogenous carbohydrate oxidation from total carbohydrate oxidation.

184

185 Computations were made on the assumption that, in response to exercise, ^{13}C is not irreversibly
186 lost in pools of tricarboxylic acid cycle intermediates and/or bicarbonate, and that lactate produced
187 from either glucose or fructose is either oxidized in muscle or recycled through gluconeogenesis
188 to be used subsequently by complete oxidation. Essentially exogenous carbohydrate oxidation is
189 calculated irrespective of the pathway that finally produces $^{13}\text{CO}_2$ that can be measured. The
190 calculations are based on the assumption that $^{13}\text{CO}_2$ recovery in expired gases was complete or
191 almost complete during exercise (31). Such computation has been shown to underestimate
192 exogenous carbohydrate oxidation rates at the beginning of exercise because of the delay between
193 $^{13}\text{CO}_2$ production in tissues and expired $^{13}\text{CO}_2$ at the mouth (32). Based on this, exogenous
194 carbohydrate oxidation rates are presented from 60 minutes onwards during the exercise period,
195 where it is expected that there would be isotopic equilibrium in the tissues and at the mouth.

196
197 Using the isotopic compositions of plasma glucose (R_{glu}) the oxidation rate of plasma glucose was
198 computed at 60, 90 and 120 minutes during exercise (equation 2 (33)).

199
200 Plasma glucose oxidation ($\text{g}\cdot\text{min}^{-1}$) = $\dot{V}\text{CO}_2 [(R_{\text{exp}} - R_{\text{ref}}) / (R_{\text{glu}} - R_{\text{ref}})] / k$ (2)

201
202 The oxidation rate of muscle glycogen ($\text{g}\cdot\text{min}^{-1}$), either directly or through the lactate shuttle (34),
203 was calculated by subtracting plasma glucose oxidation from total carbohydrate oxidation. Finally,
204 the amount of glucose released from the liver was estimated as the difference between plasma
205 glucose and exogenous carbohydrate oxidation (33).

206
207

208 *Statistical Analysis*

209 GraphPad Prism version 6.00 for Windows (GraphPad Software, La Jolla California USA) was
210 used for graph creation. Data were normally distributed (Shapiro-Wilk) and are presented as mean
211 \pm SD. Two-way repeated measures ANOVA was used to compare differences in fuel use and blood
212 related variables over time and between conditions. Post-hoc analysis was performed for any
213 significant effects using paired samples t-tests. Paired t-tests were also used to compare mean
214 differences in relative and absolute fuel use, as well as heart rate, SpO₂ and RPE between
215 conditions. This was supported where appropriate with 95% confidence intervals. Data were
216 evaluated using SPSS for Windows version 22 (Chicago, USA) with statistical significance
217 determined as $P < 0.05$. Due to participant attrition potentially affecting the power of the study,
218 we have placed greater emphasis on effect sizes (ES) in the interpretation of these data. Cohen's d
219 effect sizes were calculated, and interpreted using a threshold scale, where 0-0.2 was considered
220 to be a trivial effect, 0.2-0.6 a small effect, 0.6-1.2 a moderate effect, 1.2-2.0 a large effect, and
221 >2.0 a very large effect (35).

222

223 **Results**

224 **Total Carbohydrate and Fat Oxidation**

225 There was a *small*, but non-significant difference in total energy expenditure for 2 hours of
226 continuous cycling between HA and sea level conditions (1154.0 ± 170.5 kcal vs. 1245.3 ± 160.5
227 kcal, ES=0.54; $P=0.114$). *Large* and *moderate* effect sizes indicated lower absolute carbohydrate
228 oxidation at HA compared with sea level during the first hour, second hour and for the entire 2
229 hours of continuous cycling, though non-significant (table 1). *Small* non-significant effects were
230 observed for the relative contribution of carbohydrate to the total energy yield during the second

231 hour of exercise when comparing HA and sea level trials ($50.2 \pm 10.2\%$ vs. $54.9 \pm 6.3\%$, ES=0.46;
232 P=0.34). Alternatively, only *small* and *trivial* effect sizes for absolute fat oxidation were observed
233 during the exercise periods (table 1). *Small* non-significant effects were also observed for the
234 relative contribution of fat oxidation to the total energy yield during the second hour of exercise
235 when comparing HA and sea level trials ($49.8 \pm 10.2\%$ vs. $45.08 \pm 6.3\%$; ES =0.46; P=0.34), figure
236 1.

237 238 **Exogenous and Endogenous Carbohydrate oxidation.**

239 Exogenous carbohydrate oxidation rates were *moderately* lower at HA compared with sea level,
240 during the second hour of continuous cycling, being significant at 60 (ES=0.99, P=0.003), 90
241 (ES=1.19, P=0.017) and 120 minutes (ES=0.99, P=0.024), figure 2a. A *small* effect was observed
242 for the relative contribution of exogenous carbohydrate oxidation to the total energy yield between
243 HA and sea level, which approached significance ($32.6 \pm 6.1\%$ vs. $36.0 \pm 6.1\%$, ES=0.56,
244 P=0.059), figure 1. Further, absolute exogenous carbohydrate oxidation during the second hour of
245 exercise, was *moderately* and significantly lower at HA compared with sea level (table 2). There
246 was a *moderate* but non-significant effect towards lower absolute endogenous carbohydrate
247 oxidation at HA compared with sea level for the second hour of exercise (table 2). Furthermore,
248 the relative contribution of endogenous carbohydrate oxidation to the total energy yield was *trivial*
249 and non-significant between HA ($17.6 \pm 13.2\%$) and sea level ($19.0 \pm 9.8\%$, ES=0.1, P=0.725).

250 251 **Oxidation of plasma glucose, liver glucose and muscle glycogen**

252 A lower rate of plasma glucose oxidation was seen at HA compared with sea level during the
253 second hour of exercise, with *moderate* (60 minutes, ES=1.07) and *large* (90 minutes, ES=1.61

254 and 120 minutes, ES=1.33) but non-significant effects (P=0.113), figure 2b. In addition, the
255 absolute contribution of plasma glucose to the total energy yield was *moderately* and significantly
256 lower at HA compared with sea level (table 2). The rate of liver glucose oxidation produced non-
257 significant (P=0.471) *moderate* (60 minutes, ES=0.75), *trivial* (90 minutes, ES=0.12) and *small*
258 effects (120 minutes, ES=0.36) between conditions, figure 2c. Further, *trivial* and non-significant
259 effects were observed for both the relative (HA: $3.2 \pm 1.2\%$ vs. sea level: $3.1 \pm 0.8\%$, ES=0.09,
260 P=0.635, figure 1) and absolute contributions of liver glucose (table 2) to the total energy yield
261 between conditions during the second hour of exercise. The rate of muscle glycogen oxidation
262 produced non-significant *moderate* (60 minutes, ES=0.92, P=0.085) and *trivial* effects (90
263 minutes, ES=0.04, P=0.841 and 120 minutes, ES=0.15, P=0.708) between conditions, figure 2d.
264 *Trivial* non-significant effect was observed for the relative contribution of muscle glycogen to the
265 total energy yield for the second hour of exercise when comparing HA and sea level ($14.4 \pm 12.2\%$
266 vs. $15.8 \pm 9.3\%$, ES=0.11, P=0.934), figure 1. Furthermore, the absolute contributions of muscle
267 glycogen showed a *small* non-significant effect between conditions (table 2).

268

269 **Blood Biochemistry**

270 Plasma glucose concentrations were *moderately* higher at HA compared with sea level at 90 and
271 120 minutes (ES=0.74 and 0.87), with the condition and time interaction approaching significance
272 (P=0.072), figure 3a. *Moderate to large effect* sizes indicate higher plasma lactate concentrations
273 during exercise at HA compared with sea level (ES=0.86), but failed to reach statistical
274 significance (P=0.324), figure 3b. There were mainly *small* and non-significant effects for serum
275 insulin responses during exercise between conditions, which approached significance (ES<0.60,
276 P=0.07), figure 3c. Serum free fatty acid concentrations were *moderately* higher during the initial

277 30 minutes of exercise at HA compared with sea level (ES>0.69), but there was a non-significant
278 condition and time interaction (P=0.469), figure 3d. Metanephrine concentration was *moderately*
279 higher at HA compared with sea level at 60 minutes (ES=0.94) and *largely* higher at 120 minutes
280 (ES= 1.40), however, these differences only approached significance (P>0.075), figure 3e. A *very*
281 *large* effect for normetanephrine concentration was observed, being significantly higher at HA
282 compared with sea level at both 60 (ES= 2.02, P=0.009,) and 120 minutes (ES=2.36, P=0.006),
283 figure 3f.

284

285 **Heart Rate, Rating of Perceived Exertion and SpO₂**

286 Table 3 shows heart rate, RPE and SpO₂ at HA and sea level during the 2 hours of cycling, as well
287 as during the initial and last hour of cycling.

288

289 **Discussion**

290

291 This study, to our knowledge, is the first to compare exogenous and endogenous (liver and muscle)
292 carbohydrate oxidation, as well as fat oxidation during matched relative intensity (55% W_{max})
293 cycling at terrestrial HA and sea level in women. Exogenous carbohydrate oxidation supplied as
294 glucose and fructose made a significant contribution to the total energy yield in both conditions
295 during the second hour of exercise. However, the primary findings are that exogenous
296 carbohydrate oxidation was reduced in acute hypoxia compared with normoxia in these women
297 leading to lower absolute whole body carbohydrate oxidation at HA. The latter was not associated
298 with alterations in the use of endogenous glycogen stores.

299

300 The suppressed rate of exogenous carbohydrate oxidation at HA (0.86 g.min⁻¹) compared with sea
301 level (1.03 g.min⁻¹) is supported by literature in men when the relative (10) and absolute (29)

302 exercise intensity was matched. Our previous study in men produced similar rates of exogenous
303 carbohydrate oxidation at HA ($0.92 \text{ g}\cdot\text{min}^{-1}$) (10). In contrast to our earlier work in men (10), the
304 differences in the rates of exogenous carbohydrate oxidation in these women was large enough for
305 the relative and absolute contributions to the total energy yield to demonstrate the same pattern of
306 response, being lower at acute hypoxia compared with sea level. These differences in the absolute
307 and relative exogenous carbohydrate responses between men and women who took part in an
308 identical study remains to be explained. Young et al. (29) reported suppressed exogenous glucose
309 oxidation in men, *albeit* when the absolute exercise intensity was matched between HA (0.19
310 $\text{g}\cdot\text{min}^{-1}$) and sea level ($0.38 \text{ g}\cdot\text{min}^{-1}$). However, their rates were lower compared with the present
311 study, which in part may be due to the lower workload, as well as the different mode of exercise
312 compared with the present study (walking vs. cycling). Further, their study is limited as the
313 glucose-fructose beverage was only enriched with ^{13}C - glucose, hence their data is likely
314 underestimated as based solely on exogenous glucose oxidation, not including any fructose
315 contribution. In contrast, Peronnet et al. (9) showed no differences in exogenous glucose oxidation
316 following glucose ingestion ($1.75 \text{ g}\cdot\text{min}^{-1}$) when matching both the relative and absolute exercise
317 intensity between HA and sea level. In comparison the rates of exogenous carbohydrate oxidation
318 were also lower at both HA ($0.43 \text{ g}\cdot\text{min}^{-1}$) and sea level (relative: $0.54 \text{ g}\cdot\text{min}^{-1}$, absolute: 0.50
319 $\text{g}\cdot\text{min}^{-1}$) compared with the present study. The greater availability of carbohydrate in the present
320 study is due to the likely higher overall carbohydrate absorption rates (36), due to glucose and
321 fructose's distinctly different intestinal transport mechanisms.

322

323 Acute hypoxic exposure is associated with reduced insulin sensitivity in both men (37) and women
324 (38) due to increased catecholamine's and cortisol levels. However, overall the hyperglycemia in

325 both conditions was similar, which is in contrast to studies that have reported elevated plasma
326 glucose concentrations following carbohydrate ingestion during acute hypoxia compared with sea
327 level (9, 29). Furthermore, the plasma glucose concentrations in the present study, stimulated
328 insulin to similar concentrations, despite at least *moderately* higher metanephrine and
329 normetanephrine concentrations at HA. Therefore, the present study does not support the idea of
330 insulin resistance being a plausible explanation for the reduced exogenous carbohydrate oxidation
331 during exercise on acute exposure to HA.

332
333 The absolute whole body carbohydrate oxidation was reduced in women at HA compared with sea
334 level. These data are consistent with the previous literature in men (9, 10), following glucose-
335 fructose or glucose ingestion during matched relative exercise intensity. When comparing these
336 data with our previous research in men (10), the reduction in whole body carbohydrate oxidation
337 during 2 hours of exercise at HA is less (~40g vs. ~129g). This may be explained by the lower
338 absolute cycling intensity for the women (105.0 ± 17.2 watts) compared with the men (114.9 ± 9.7
339 watts), with a lower total energy expenditure (~1154 kcal vs. ~1347 kcal). However, preferential
340 use of fat as a fuel source by women may also be an explanation.

341 A small difference of 7% in total energy expenditure between HA and sea level is due to the lower
342 absolute workload at HA (105.0 ± 17.2 vs. 114.7 ± 18.9 watts). This will have only marginally
343 contributed to the lower absolute oxidation of whole body carbohydrate at HA and is unlikely to
344 be the full explanation as the magnitude of difference (~24%) far outweighs any difference in total
345 energy expenditure. The primary explanation is that reduced exogenous carbohydrate oxidation
346 makes the most significantly contribution towards the lower whole body carbohydrate oxidation
347 during the second hour of exercise at HA.

348 In contrast to the literature in men (10, 39), absolute and relative fat oxidation did not significantly
349 increase during exercise at HA compared with sea level. These data are supported by the similar
350 free fatty acid concentrations, suggesting the rate of utilization is comparable despite the higher
351 normetanephrine and metanephrine concentrations. The fact that fat oxidation was not
352 significantly affected by acute exposure to HA in these women, may be due to their higher fat
353 oxidation at sea level (baseline effect) compared with men. This has been ascribed to women
354 having a greater proportion of body fat (12), a greater lipid content in muscles fibers (11), a higher
355 percentage of type 1 muscle fibers (40) and better mobilization of fatty acids from subcutaneous
356 adipose tissue linked to cyclic changes in estrogen and progesterone (13). However, it is interesting
357 to note that the magnitude of fat oxidation in women at HA, was very similar compared with our
358 previous study in men (50% vs. 51%), as was the use of pre-existing muscle glycogen (17% vs.
359 16%), exogenous carbohydrate oxidation (32% vs. 30%) and glucose release from the liver (3%).
360 This suggests that women and men fuel exercise at HA in a very similar fashion following the
361 ingestion of glucose-fructose.

362
363 The sample size in the present study is smaller than originally intended, due to participant attrition.
364 Post-hoc power analysis revealed that the rate of exogenous carbohydrate oxidation provided a
365 power of 57%, 71% and 57% for 60, 90 and 120 minutes of cycling, respectively, using the mean
366 difference and associated standard deviation at each time point and an alpha level of 0.05.
367 Therefore, this study is slightly underpower compared with the accepted 80% and should be
368 interpreted in this context.

369
370 In conclusion, acute exposure to hypoxia reduced the exogenous oxidation of glucose and fructose
371 compared with normoxia during 2 hours of cycling at the same relative exercise intensity. This

372 may have practical implications, as the conventional carbohydrate ingestion recommendations
373 alter fuel use compared with normoxia. Thus, further research is required to identify an optimal
374 dose for women on acute HA exposure, as well as establishing the mechanism for reduced
375 exogenous carbohydrate oxidation.

376

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383 inappropriate data manipulation. The results of the present study do not constitute endorsement by
384 ACSM.

385

386 **Conflict of Interest**

387 The authors report no conflict of interest.

388

389 **References**

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391 1. Hochachka PW, Stanley C, Matheson GO, McKenzie DC, Allen PS, Parkhouse WS.

392 Metabolic and work efficiencies during exercise in Andean natives. *J Appl Physiol* (1985).

393 1991;70(4):1720-30.

394 2. Griffiths A, Shannon OM, Matu J, King R, Deighton K, O'Hara JP. The effects of

395 environmental hypoxia on substrate utilisation during exercise: a meta-analysis. *J Int Soc Sports*

396 *Nutr.* 2019;16(1):10. Epub 2019/03/01.

- 397 3. Brooks GA, Butterfield GE, Wolfe RR, et al. Increased dependence on blood glucose after
398 acclimatization to 4,300 m. *J Appl Physiol* (1985). 1991;70(2):919-27.
- 399 4. Roberts AC, Reeves JT, Butterfield GE, et al. Altitude and beta-blockade augment glucose
400 utilization during submaximal exercise. *J Appl Physiol* (1985). 1996;80(2):605-15.
- 401 5. Katayama K, Goto K, Ishida K, Ogita F. Substrate utilization during exercise and recovery
402 at moderate altitude. *Metabolism*. 2010;59(7):959-66.
- 403 6. Friedmann B, Bauer T, Menold E, Bartsch P. Exercise with the intensity of the individual
404 anaerobic threshold in acute hypoxia. *Med Sci Sports Exerc*. 2004;36(10):1737-42.
- 405 7. Young AJ, Evans WJ, Cymerman A, Pandolf KB, Knapik JJ, Maher JT. Sparing effect of
406 chronic high-altitude exposure on muscle glycogen utilization. *Journal of applied physiology:
407 respiratory, environmental and exercise physiology*. 1982;52(4):857-62.
- 408 8. Bouissou P, Guezennec CY, Defer G, Pesquies P. Oxygen consumption, lactate
409 accumulation, and sympathetic response during prolonged exercise under hypoxia. *Int J Sports
410 Med*. 1987;8(4):266-9. Epub 1987/08/01.
- 411 9. Peronnet F, Massicotte D, Folch N, et al. Substrate utilization during prolonged exercise
412 with ingestion of (13)C-glucose in acute hypobaric hypoxia (4,300 m). *Eur J Appl Physiol*.
413 2006;97(5):527-34.
- 414 10. O'Hara JP, Woods DR, Mellor A, et al. A comparison of substrate oxidation during
415 prolonged exercise in men at terrestrial altitude and normobaric normoxia following the
416 coingestion of 13C glucose and 13C fructose. *Physiological reports*. 2017;5(1).
- 417 11. Tarnopolsky MA. Gender differences in substrate metabolism during endurance exercise.
418 *Canadian journal of applied physiology = Revue canadienne de physiologie appliquee*.
419 2000;25(4):312-27.

- 420 12. Knechtle B, Muller G, Willmann F, Kotteck K, Eser P, Knecht H. Fat oxidation in men
421 and women endurance athletes in running and cycling. *Int J Sports Med.* 2004;25(1):38-44. Epub
422 2004/01/30. doi: 10.1055/s-2003-45232.
- 423 13. Frontera WR, Suh D, Krivickas LS, Hughes VA, Goldstein R, Roubenoff R. Skeletal
424 muscle fiber quality in older men and women. *American journal of physiology Cell physiology.*
425 2000;279(3):C611-8.
- 426 14. Tremblay J, Peronnet F, Massicotte D, Lavoie C. Carbohydrate supplementation and sex
427 differences in fuel selection during exercise. *Med Sci Sports Exerc.* 2010;42(7):1314-23.
- 428 15. M'Kaouar H, Peronnet F, Massicotte D, Lavoie C. Gender difference in the metabolic
429 response to prolonged exercise with [13C]glucose ingestion. *Eur J Appl Physiol.* 2004;92(4-
430 5):462-9.
- 431 16. Riddell MC, Partington SL, Stupka N, Armstrong D, Rennie C, Tarnopolsky MA.
432 Substrate utilization during exercise performed with and without glucose ingestion in female and
433 male endurance trained athletes. *Int J Sport Nutr Exerc Metab.* 2003;13(4):407-21.
- 434 17. Wallis GA, Dawson R, Achten J, Webber J, Jeukendrup AE. Metabolic response to
435 carbohydrate ingestion during exercise in males and females. *Am J Physiol Endocrinol Metab.*
436 2006;290(4):E708-15.
- 437 18. Beidleman BA, Rock PB, Muza SR, et al. Substrate oxidation is altered in women during
438 exercise upon acute altitude exposure. *Med Sci Sports Exerc.* 2002;34(3):430-7.
- 439 19. Lundby C, Van Hall G. Substrate utilization in sea level residents during exercise in acute
440 hypoxia and after 4 weeks of acclimatization to 4100 m. *Acta physiologica Scandinavica.*
441 2002;176(3):195-201.

- 442 20. Kuipers H, Keizer HA, Brouns F, Saris WH. Carbohydrate feeding and glycogen synthesis
443 during exercise in man. *Pflugers Arch.* 1987;410(6):652-6.
- 444 21. Conkin J. PH₂O and simulated hypobaric hypoxia. *Aviation, space, and environmental*
445 *medicine.* 2011;82(12):1157-8.
- 446 22. Wasserman K, Hansen J, Sue D, Casaburi R, Whipp B. Principles of exercise testing and
447 interpretation: including pathophysiology and clinical applications. 3rd ed. Philadelphia:
448 Lippincott Williams & Wilkins; 1999. p64-68
- 449 23. Morrison DJ, O'Hara JP, King RF, Preston T. Quantitation of plasma ¹³C-galactose and
450 ¹³C-glucose during exercise by liquid chromatography/isotope ratio mass spectrometry. *Rapid*
451 *Commun Mass Spectrom.* 2011;25(17):2484-8.
- 452 24. Craig H. Isotopic standards for carbon and oxygen and correction factors for mass-
453 spectrometric analysis of carbon dioxide. *Geochimica et Cosmochimica Acta.* 1957;12(1-2):133-
454 49.
- 455 25. Peronnet F, Massicotte D. Table of nonprotein respiratory quotient: an update. *Can J Sport*
456 *Sci.* 1991;16(1):23-9.
- 457 26. Craig H. The geochemistry of the stable carbon isotopes. *Geochim Cosmochim Acta.*
458 1953;3:53-92.
- 459 27. Mosora F, Lacroix M, Luyckx A, et al. Glucose oxidation in relation to the size of the oral
460 glucose loading dose. *Metabolism.* 1981;30(12):1143-9.
- 461 28. Peronnet F, Massicotte D, Brisson G, Hillaire-Marcel C. Use of ¹³C substrates for
462 metabolic studies in exercise: methodological considerations. *J Appl Physiol.* 1990;69(3):1047-
463 52.

- 464 29. Young AJ, Berryman CE, Kenefick RW, et al. Altitude Acclimatization Alleviates the
465 Hypoxia-Induced Suppression of Exogenous Glucose Oxidation During Steady-State Aerobic
466 Exercise. *Frontiers in physiology*. 2018;9:830.
- 467 30. Gautier JF, Pirnay F, Jandrain B, et al. Endogenous substrate oxidation during exercise and
468 variations in breath $^{13}\text{CO}_2/^{12}\text{CO}_2$. *J Appl Physiol* (1985). 1993;74(1):133-8.
- 469 31. Trimmer JK, Casazza GA, Horning MA, Brooks GA. Recovery of $(^{13}\text{C})\text{CO}_2$ during rest
470 and exercise after $[1-(^{13}\text{C})\text{acetate}]$, $[2-(^{13}\text{C})\text{acetate}]$, and $\text{NaH}(^{13}\text{C})\text{CO}_3$ infusions. *Am J Physiol*
471 *Endocrinol Metab*. 2001;281(4):E683-92.
- 472 32. Pallikarakis N, Sphiris N, Lefebvre P. Influence of the bicarbonate pool and on the
473 occurrence of $^{13}\text{CO}_2$ in exhaled air. *Eur J Appl Physiol Occup Physiol*. 1991;63(3-4):179-83.
- 474 33. Peronnet F, Rheume N, Lavoie C, Hillaire-Marcel C, Massicotte D. Oral $[^{13}\text{C}]\text{glucose}$
475 oxidation during prolonged exercise after high- and low-carbohydrate diets. *J Appl Physiol*.
476 1998;85(2):723-30.
- 477 34. Brooks GA. Lactate production under fully aerobic conditions: the lactate shuttle during
478 rest and exercise. *Federation proceedings*. 1986;45(13):2924-9.
- 479 35. Batterham AM, Hopkins WG. Making meaningful inferences about magnitudes.
480 *International journal of sports physiology and performance*. 2006;1(1):50-7.
- 481 36. Shi X, Summers RW, Schedl HP, Flanagan SW, Chang R, Gisolfi CV. Effects of
482 carbohydrate type and concentration and solution osmolality on water absorption. *Med Sci Sports*
483 *Exerc*. 1995;27(12):1607-15.
- 484 37. Larsen JJ, Hansen JM, Olsen NV, Galbo H, Dela F. The effect of altitude hypoxia on
485 glucose homeostasis in men. *The Journal of physiology*. 1997;504 (Pt 1):241-9.

486 38. Braun B, Horton T. Endocrine regulation of exercise substrate utilization in women
487 compared to men. *Exercise and sport sciences reviews*. 2001;29(4):149-54.

488 39. Matu J, Deighton K, Ispoglou T, Duckworth L. The effect of moderate versus severe
489 simulated altitude on appetite, gut hormones, energy intake and substrate oxidation in men.
490 *Appetite*. 2017;113:284-92.

491 40. Glenmark B. Skeletal muscle fibre types, physical performance, physical activity and
492 attitude to physical activity in women and men. A follow-up from age 16 to 27. *Acta Physiol Scand*
493 *Suppl*. 1994;623:1-47.

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Figure Captions

Figure 1. The relative (% of energy yield) contribution of exogenous and endogenous substrate oxidation during the second hour of cycling at high altitude and sea level.

Figure 2. Oxidation rates of exogenous CHO (a), plasma glucose (b), glucose released from the liver (c) and muscle glycogen (d) during the second hour of cycling. * high altitude significantly lower compared with sea level ($P<0.05$).

Figure 3. Plasma glucose (a), plasma lactate (b), serum insulin (c), serum free fatty acids (d), plasma metanephrine (e) and plasma normetanephrine (f) concentrations at rest and during 2 hours of cycling. * high altitude significantly higher compared with sea level ($P<0.05$).

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