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Echocardiographic changes following Active Heat Acclimation

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Key Words

heat adaptation; heat acclimation; plasma volume; stroke volume; diastolic function; echocardiography; preload

Abstract

Heat adaption through acclimatisation or acclimation improves cardiovascular stability by maintaining cardiac output due to compensatory increases in stroke volume. The main aim of this study was to assess whether 2D transthoracic echocardiography (TTE) could be used to confirm differences in resting echocardiographic parameters, before and after active heat acclimation (HA). Thirteen male endurance trained cyclists underwent a resting blinded TTE before and after randomisation to either 5 consecutive daily exertional heat exposures of controlled hyperthermia at 32°C with 70% relative humidity (RH) (HOT) or 5-days of exercise in temperate (21°C with 36% RH) environmental conditions (TEMP). Measures of HA included heart rate, gastrointestinal temperature, skin temperature, sweat loss, total nonurinary fluid loss (TNUFL), plasma volume and participant's ratings of perceived exertion (RPE). Following HA, the HOT group demonstrated increased sweat loss (p=0.01) and TNUFL (p=0.01) in comparison to the TEMP group with a significantly decreased RPE (p=0.01). On TTE, post exposure, there was a significant comparative increase in the HOT group in left ventricular end diastolic volume (p=0.029), SV (p=0.009), left atrial volume (p=0.005), inferior vena cava diameter (p=0.041), and a significant difference in mean peak diastolic mitral annular velocity (e') (p=0.044). Cardiovascular adaptations to HA appear to be

predominantly mediated by improvements in increased preload and ventricular compliance.

TTE is a useful tool to demonstrate and quantify cardiac HA.

Introduction

Thermoregulatory induced cardiovascular insufficiency impairs the ability to exercise in the heat^{1,2}. Heat adaption through acclimatisation or acclimation (HA) improves cardiovascular stability by maintaining cardiac output, despite lowering resting heart rate (HR)³, due to compensatory increases in stroke volume (SV)^{1,4}. The increase in SV is predominantly thought to be due to an increase in plasma volume (PV)^{5,6} however the effect size detected by meta-analysis is small¹. Improvements in cardiovascular efficiency following HA have also been attributed to falls in the core and skin temperature accommodating a lower HR and consequent increased SV⁷. Other mechanisms include a reduction in sympathetic stimulation⁸ which has been shown to reduce HR and induce plasma volume expansion in animal models⁹. Fundamental changes in the cardiac morphology have also been identified, following HA, with increased ventricular compliance^{9,10} secondary to a redistribution of cardiac myosin isoenzymes and alterations in sarcoplasmic proteins in rat models^{11,12}. This allows the ventricle to accept an increased preload and then to deliver an increased SV without an increase in end-diastolic pressure. This increase in compliance improves cardiac efficiency and pressure generation^{10,13} so advantageously increasing cardiac output without a significant increase in HR. This suite of cardiovascular changes has not been conclusively demonstrated in human subjects.

This study was performed to specifically assess whether resting 2D trans-thoracic echocardiography (TTE) could be used to identify differences in measurable echocardiographic cardiovascular parameters following active HA in comparison to exercise in temperate conditions. A secondary aim was to investigate whether TTE could provide further insight into cardiovascular adaptations to heat and the underlying mechanisms in human subjects.

Methods

Participants

The study protocol was approved by Leeds Beckett University Research and Ethics Committee. Thirteen healthy male participants volunteered and gave written consent. All participants were amateur endurance trained cyclists, or triathletes, classified as performance level 3 by training or $\dot{V}O_{2peak}^{14}$. Each participant was previously unacclimated to the heat and the acclimation sessions occurred during the winter-spring to reduce seasonal acclimatisation effects¹⁵. Additionally, participants were asked to avoid prolonged thermal exposures¹⁶ (i.e. baths, saunas and steam rooms), during the trial period. Participants were instructed to maintain their normal training routines in the interval between initial measures and the intervention. Participants were requested not to undergo any training during the intervention period. All participants were asked to refrain from alcohol consumption at least 24 hours prior to any testing and to abstain from caffeine/stimulant containing drinks 6 hours prior to TTE.

Preliminary testing

Body composition was assessed by calibrated air displacement plethysmography (BOD POD, Life measurement systems, USA), and measurements of height (Seca, 220, Germany) and body mass (Seca, 770, Germany) were recorded. $\dot{V}O_{2peak}$ (L.min⁻¹) was determined from an incremental test to volitional exhaustion on a cycle ergometer (Wattbike, Trainer, UK) in temperate laboratory conditions (19.3 ± 1.6°C, 47± 6% relative humidity). Participants adjusted bike dimensions at baseline, and these then remained fixed for all experimental trials. Participants started cycling at ~150 watts, and increased the workload at a rate of 20 W.min⁻¹ until exhaustion or the individual's cadence dropped below 70 rpm¹⁷. Respiratory gases were measured continuously using breath-by-breath in-line gas analysis (Cortex, Metalyser, 3B, Germany), calibrated following the manufacturer's instructions. HR was recorded continuously during all exercise tests (Polar, V800, Finland). \dot{VO}_{2peak} was considered the highest \dot{VO}_2 in any 30s period in the last two minutes¹⁸.

Echocardiography

Prior to intervention participants underwent a resting TTE (CX50, Phillips, USA) performed by a blinded British Society of Echocardiography (BSE) accredited practitioner. Echocardiography measures were performed in the resting partial left decubitus position with the couch head at 30° elevation (flat for subcostal views) satisfying the requirements of the BSE minimum data set¹⁹. Measures were averaged over 3 heart beats. TTE was performed at a similar time of day to avoid potential diurnal variation. Left ventricular stroke volume (SV; in cm³) was calculated in two ways: 1) from the product of the velocity-time integral (cm) of the pulsed-wave Doppler in the left ventricular outflow tract (LVOT) and the LVOT cross sectional area (πr^2 ; in cm²), determined by a TTE measurement of the LVOT in the parasternal long-axis view; 2) From the subtraction of the left ventricular end systolic volume (LVESV) from the left ventricular end diastolic volume (LVEDV) calculated using Simpson biplane method¹⁹ in both two chamber and four chamber views, and then averaged. Atrial volumes were calculated using the area-length method²⁰. The septal and lateral mitral annular peak velocities were averaged to give an overall e'.

Acclimation protocol

Participants were randomised to either an isothermic HA (HOT) (n=7) or temperate exercise intervention (TEMP) (n=6). The isothermic HA arm consisted of five consecutive daily exertional heat exposures performed in an environmental chamber at $32.1 \pm 0.1^{\circ}$ C with 70.3 \pm 1.1% relative humidity, utilising the controlled hyperthermia method which is potentially more effective in eliciting greater and more rapid HA than constant work or self-paced approaches^{5,21}. This approach involves rapidly increasing core temperature (1-1.5°C) through an active exercise phase, followed by a reduced exercise intensity to attain and maintain a core temperature (\geq 38.5°C). This was achieved by cycling at a relative exercise intensity of 2.0-2.7 W ·kg⁻¹ ²² to reach the target temperature in ~30-minutes and maintain it for 60 minutes with the work titrated to maintain this core temp. The TEMP group cycled for 90 minutes, every day for 5 days in temperate environmental conditions (21.5±1.2°C with 36.2± 5.6% relative humidity) at 2.0-2.7 W ·kg⁻¹.

Two hours before arrival at each experimental exposure, participants consumed 500mL of water to ensure euhydration prior to each exposure. On arrival, participants provided a urine sample which was assessed for urine colour, osmolality (Osmocheck, Vitech, Scientific Ltd, Japan) and specific gravity (Hand refractometer, Atago, Tokyo, Japan). Participants were deemed euhydrated when urine osmolality was <700 mOsmol \cdot kg⁻¹, specific gravity <1.020 and urine colour <3 on an 8-point scale²³.

Prior to commencing exercise, participants' nude body mass (NBM) was assessed and haemoglobin (Hb) concentration and haematocrit (Hct) measured, in triplicate, to calculate the plasma volume (PV) using the Dill and Costill method²⁴. This was followed by 5-minutes rest in HOT or TEMP conditions, after which resting HR, gastrointestinal temperature (T_{gi}), skin temperature (T_{sk}), thermal sensation score (TSS)²⁵ and thermal comfort²⁶ (TC) were recorded and the physiological strain index (PSI) was calculated²⁷. T_{gi} was measured by using a calibrated²⁸ ingestible core temperature pill (BodyCap, e-Celsius, France), set to record at 10s intervals, ingested >6 hours before the trial. Four skin temperature loggers (iButton TM, Integrated Products, USA) were attached to the right side of the body and mean skin temperature calculated from the equation by Ramanathan, 1964²⁹. HR was monitored continuously whilst TSS, RPE, TC, Tgi, T_{sk} were recorded at 5-minute intervals. NBM was measured post exposure and whole body sweat rate and total non-urinary fluid loss were calculated. Fluid intake was consumed *ad libitum* by participants and weighed to the nearest (g) to calculate whole body sweat rate ((preNBM – postNBM) + (fluid intake - urine output)/time). Following completion of the five-day acclimation/control the TTE, and PV was re-assessed within 24 hours.

Statistical Analysis

Measures were assessed for normality using the Shapiro–Wilk test prior to data analysis. Physical characteristics of the two groups were compared using an unpaired t-test. Physiological measures were compared between the HOT and TEMP groups on Day 1 and Day 5 using a two-way ANOVA. Nominal categorical variables were measured using a Kruskal-Wallis ANOVA on the Δ change (Day 5- Day 1) between HOT and TEMP.

The difference in TTE measures (post exposure-pre exposure) was calculated and compared between HOT and TEMP groups. Cohen's d effect size (ES) was calculated, together with 95% confidence intervals, using a threshold scale where an ES of 0-0.2 was considered trivial; 0.2-0.6, a *small* effect; 0.6-1.2, a *moderate* effect; 1.2-2.0, a *large* effect; and >2, a *very large* effect³⁰. A 2-way ANOVA of TTE parameters was performed using HOT and TEMP and pre and post exposure. ES and CI were also calculated for the Δ change in PV between HOT and TEMP.

The α level was set to 0.05. All statistical analyses were performed using GraphPad Prism 8.0, GraphPad Software, San Diego, California.

Results

All participants completed the exposures in their entirety. The mean age, body surface area (Mosteller) and $\dot{V}O_{2peak}$ was 28.5 (±9.9), 1.93±0.1 m² and 59.7±8.1ml·kg·min⁻¹ respectively.

There were no significant differences in age, height, body mass, body surface area, body fat percentage, or $\dot{V}O_{2peak}$ between HOT and TEMP groups.

There was a (9±12%) increase in PV in the HOT group with a -0.2±6% increase in the TEMP with a *moderate*, but insignificant, trend towards a Δ increase in PV ES 1.02 (CI -0.19 to 2.24) (p=0.102) as assessed by the Dill and Costill method²⁴. Following HA, the HOT group demonstrated increased sweat loss, from day 1 to day 5 (0.36±0.28 L/min), in comparison to the TEMP group (-0.07±0.19L/min) (p=0.01) with Δ increases in total non-urinary fluid loss (TNUFL) (HOT; 548ml±430ml, TEMP; -110ml±291ml, p=0.01). There was no significant change, from day 1 to day 5, in resting heart rate (HOT; -5±4.6bpm, TEMP; 1±7.4bpm) or peak heart rate (HOT; -4.9±6.2bpm, TEMP; -2±9.4bpm) between groups. There was a significantly decreased RPE (p=0.01) in the HOT group (-2±1)in comparison to the TEMP group (0±1) but no significant difference in the T_{gl}, T_{sk}, TSS, TC or PSI. With regard to hydration status, there was no significant difference in urine osmolality, urine colour, urine specific gravity or body mass in comparing day 1 and day 5 exposure values between HOT and TEMP groups.

TTE data are detailed in Table 1. Pre-exposure, indexed left ventricular volumes (LVEDV HOT 80±7ml/m², TEMP 80±11ml/m²) and atrial volumes (LA HOT 30±6ml/m², TEMP 31±11ml/m²) were elevated in both groups in comparison to normal reference values¹⁹. There were no significant differences between the HOT and TEMP groups in any of the baseline TTE variables. The change in significant echocardiographic variables (post exposure -pre exposure), comparing the HOT and TEMP group, for LA Volume, SV, e', LVEDV and IVC diameter can be visualised in Figure 1.

Discussion

The significant positive findings of this study are that a relatively short 5-day isothermic HA protocol, compared to equivalent temperate exercise, increases LA volume, SV, LVEDV, e' and IVC diameter. The LA and LV volumes were elevated in both groups at baseline, consistent with trained amateur athletes at performance Level 3. In this context, it is striking that the left atrial volumes and SV rose further following active HA in the HOT group; an effect that was not seen in the TEMP group who underwent matched exercise exposure in temperate conditions. The significant differences in thermal comfort, sweat loss and TNUFL following the active HA in the HOT group suggest at least partial acclimation.

The significant increase in LA volume and IVC diameter would suggest an increase in preload secondary to increased PV⁶. PV expansion is also likely to be at least partially responsible for the elevated SV^{1.6,31}. However, despite a *moderate* effect size, the trend towards increased PV in the HOT group was insignificant when assessed by the Dill and Costill method²⁴. This apparent contradiction might be explained by the small sample size as well as inaccurate methodology where the haemoconcentration effect is estimated by calculations based on haemoglobin concentration and haematocrit before and after exposure using the Dill and Costill method^{32–35}. Alternatively, trained individuals, such as in this study, may also have a lower PV adaptation response as measured by this methodology due to higher levels of background adaption secondary to the physical training³⁶ so requiring a longer acclimation protocol, and more participants, to exhibit the full phenotype.

PV expansion alone may not account for all the observed TTE data. The rise in the speed of early LV relaxation (e') during diastole reflects increased ventricular compliance or reduced LV stiffness. Increased compliance permits the heart to convert an elevated venous return, and consequently increased LV preload, into a higher end-diastolic volume, without elevation of LV end-diastolic pressure (LVEDP). It is known that e', the average of septal and lateral peak mitral annular velocity during early LV relaxation, increases following long-term physical training³⁷ and increases further during exercise³⁸. However e' transiently decreases immediately following high-volume exercise³⁹. HA appeared to protect against this effect when comparing e' between HOT and TEMP groups. In rats, HA redistributes cardiac myosin isoenzymes from the fast V₁ isoform with high ATPase activity to the predominance of the slow V₃ form with low ATPase activity resulting in increased contractile efficiency⁹ and increased LV compliance. This is not thought to be due to volume increases but a true change in heat-acclimated myocardial elastic properties⁹. The combined heat exposure in the HOT group (450 minutes) was significantly less than the stimulus in rodent studies^{10,11} where intrinsic myocardial effects were demonstrated and arguably insufficient to alter myocardial elastic properties. However a high level of physical fitness is thought to improve the physiological response to exercise in the heat⁴⁰, perhaps by 'priming', leading to a more rapid acclimation³⁶ although possibly a lower magnitude of response²⁶ following full HA.

Furthermore e' is a relatively, but not absolutely, load (preload and afterload) independent measure of LV relaxation⁴¹. There was a non-significant (p=0.099) *moderate* increase in E (rapid early diastolic filling) in the HOT group post-exposure which was mediated, in part, by a reduction in E (post-exposure) in the TEMP group. Albeit non-significant, the increase in Ewave velocity following HOT exposure and the relative fall in E-wave following TEMP exposure appear to reflect the LV filling effects of increased and decreased PV respectively. These parallel changes in E and e' within the HOT group explain the minimal change in their ratio: E/e'. E/e' is commonly recognised as a surrogate of LV filling pressure and a predictor of LA pressure⁴². This would indicate that following isothermic HA, the increased preload, potentially mediated by an increased PV, was balanced by an increase in LV compliance so as to permit increased LV filling without an increase in LA pressure⁴³. This balanced response is mirrored by the behaviour of the left ventricle during exercise. In the healthy heart, the increase in velocity of early mitral inflow (E) during exercise, is balanced by the energy-requiring increase in the velocity of early ventricular relaxation (e'). The overall effect is that the ratio E/e' is maintained, or even falls slightly, during exertion. This reflects a maintenance of (or slight reduction in) LA pressure, in stark contrast to the rise in E/e' seen in the failing ventricle.

It is known that HA leads to relative bradycardia⁷. It is not possible, in the current study, to distinguish whether heat-induced bradycardia, resulting in prolonged diastole has enhanced filling, so as to augment SV, or whether a combination of enhanced LV preload and contractility have augmented SV sufficiently to permit the attainment of the body's cardiac output at a lower resting heart rate. Regardless of the relative contribution of these two mechanisms, it is interesting to observe the trend (p=0.056) toward a lower peak HR in the HOT group in comparison to the TEMP condition. Heat-mediated decrease in sympathetic outflow⁵ is another potential mechanism for the augmentation of SV, via enhanced cardiac compliance.

Whilst changes of increased LA volume, LVEDV, SV and increased e' seen in the HOT group may all be seen following exercise training^{37,44} we believe there are a number of reasons that a HA effect is a much more compelling explanation. First, the TEMP group underwent a very similar training stimulus over the 5-day intervention and they did not exhibit the same augmented chamber volumes or improved diastolic function. Second, all participants were performance Level 3 athletes and exhibited a degree of cardiac adaptation to exercise, which did not differ betwen groups. Finally, the speed of adaptation is more in keeping with HA than training adaptation, which can take up to a year⁴⁴.

Overall, these data demonstrate several concurrent cardiovascular adaptations occurring in response to HA. These findings are supported by the only other study, to our knowledge, to assess echocardiographic responses to heat exposure. These include a significantly increased LA volume, LVEDV and LV lateral e' following passive heat exposure in 12 male participants without a control group or markers of HA⁴⁵. A further study assessed the effects of 14 - 21 days of passive HA with mild exercise routines in 8 patients prior to bypass surgery in comparison to ambient non-exercising controls. Here there was less diastolic dysfunction- by generated diastolic pressure curves in comparison to LV area- following surgery in the HA group in comparison to controls.

There are several limitations to this study which mean that whilst these data is hypothesis forming drawing firm conclusions on the observed changes on echocardiographic parameters is not possible. Our sample size was small (and curtailed by COVID-19 pandemic) and so likely lacked statistical power to show statistically significant changes in PV as well as HR (both peak and resting). Due to the limited prior studies assessing echocardiography in heat acclimation no formal power calculations were performed prior, however similar changes were observed in twelve participants without a control group with less robust passive heat stimulus⁴⁵. Furthermore echocardiography has significant interobserver and intraobserver variability particularly in the measurement of LV volumes⁴⁶. The measurement of SV by doppler assessment of the LVOT can be compromised by technical factors such as angular acuity of aortic blood flow, and/or off-axis aortic annular dimensions but has been shown to be reproducible. Studies have shown reasonable reproducibility with respect to diastolic function however⁴⁷.

A longer acclimation protocol may have more effectively demonstrated the underlying mechanisms. We chose a deliberately short but stressful humid isothermic HA exposure in

trained individuals which we considered might induce more rapid HA, particularly when incorporated with physical exercise^{48–50}, in comparison to passive heat exposure^{1,51–53}. Furthermore the cardiac adaptations to HA are thought to manifest in the first 4-7 days⁵. Whilst we instructed participants to continue normal training patterns before initial measures, and not to exercise during the interventions, we did not record their exercise activity outside of the study nor ensure that there was no significant changes between groups which potentially could have introduced bias- particularly if the TEMP group continued to exercise. This may explain the trend in reductions in echocardiographic parameters in the TEMP group (Figure 1, Table 1).

Conclusion

This study is the first to show differences in TTE parameters following active HA and the first to control for a training effect with a temperate exercise arm. It also gives further insight into the cardiovascular adaptations to HA which are characterised by an increase in both pre-load and ventricular compliance as demonstrated by elevated LV chamber volumes, increased SV and an increase in e', with no concomitant change in E/e'. TTE is a useful tool to demonstrate and quantify cardiac adaptation. This has several implications to utilise TTE as a research tool in elite athletes^{1,5} firefighters, miners, the military or aid workers⁵⁴ particularly in the development of tailored rapid HA regimes. Echocardiography in heat acclimation merits further study.

Conflict of Interest

The authors declare no conflict of interest.

References

- 1. Tyler CJ, Reeve T, Hodges GJ, Cheung SS. The Effects of Heat Adaptation on Physiology, Perception and Exercise Performance in the Heat: A Meta-Analysis. *Sport Med*. 2016;46(11):1699-1724. doi:10.1007/s40279-016-0538-5
- 2. Périard JD, Cramer MN, Chapman PG, Caillaud C, Thompson MW. Cardiovascular strain impairs prolonged self-paced exercise in the heat. *Exp Physiol*. 2011;96(2):134-144. doi:10.1113/expphysiol.2010.054213
- Sawka MN, Leon LR, Montain SJ, Sonna LA. Integrated physiological mechanisms of exercise performance, adaptation, and maladaptation to heat stress. *Compr Physiol*. 2011;1(4):1883-1928. doi:10.1002/cphy.c100082
- 4. Taylor NAS. Human heat adaptation. *Compr Physiol*. 2014;4(1):325-365. doi:10.1002/cphy.c130022
- 5. Périard JD, Travers GJSS, Racinais SS, et al. Cardiovascular adaptations supporting human exercise-heat acclimation. *Auton Neurosci Basic Clin*. 2016;196(2016):52-62. doi:10.1016/j.autneu.2016.02.002
- 6. Senay LCJ. An inquiry into the role of cardiac filling pressure in acclimatization to heat. *Yale J Biol Med.* 1986;59(3):247-256.
- 7. Rowell LB, Kraning KK, Kennedy JW, Evans TO. Central circulatory responses to work in dry heat before and after acclimatization. *J Appl Physiol*. 1967;22(3):509-518.
- Stacey MJMMJ, Delves SK, Woods DR, et al. Heart rate variability and plasma nephrines in the evaluation of heat acclimatisation status. *Eur J Appl Physiol*. 2018;118(1):165-174. doi:10.1007/s00421-017-3758-y
- 9. Horowitz M. Matching the heart to heat-induced circulatory load: heat-acclimatory responses. *News Physiol Sci an Int J Physiol Prod jointly by Int Union Physiol Sci Am Physiol Soc.* 2003;18(15):215-221.
- 10. Horowitz M, Shimoni Y, Parnes S, Gotsman MS, Hasin Y. Heat acclimation: cardiac performance of isolated rat heart. *J Appl Physiol*. 1986;60(1):9-13. doi:10.1152/jappl.1986.60.1.9
- 11. Horowitz M, Peyser YM, Muhlrad A. Alterations in Cardiac Myosin Isoenzymes Distribution as an Adaptation to Chronic Environmental Heat Stress in the Rat. *J Mol Cell Cardiol*. 1986;18:511-515.
- 12. Kiss E, Jakab G, Kranias EG, Edes I. Thyroid Hormone-Induced Alterations in Phospholamban Protein Expression Regulatory Effects on Sarcoplasmic Reticulum Ca2 ' Transport and Myocardial Relaxation. *Circ Res.* 1994;75(2):245-251.
- 13. Horowitz M. From molecular and cellular to integrative heat defense during exposure to chronic heat. *Comp Biochem Physiol A Mol Integr Physiol*. 2002;131(3):475-483. doi:10.1016/S1095-6433(01)00500-1
- De Pauw K, Roelands B, Cheung SS, de Geus B, Rietjens G, Meeusen R. Guidelines to classify subject groups in sport-science research. *Int J Sports Physiol Perform*. 2013;8(2):111-122.
- 15. Shapiro Y, RW H, Kimbrough CM, et al. Physiological and hematologic responses to summer and winter dry-heat acclimation. *J Appl Physiol*. 1981;50(4):792-798. doi:10.1152/jappl.1981.50.4.792
- 16. Stanley J, Halliday A, D'Auria S, Buchheit M, Leicht AS. Effect of sauna-based heat

acclimation on plasma volume and heart rate variability. *Eur J Appl Physiol*. 2015;115(4):785-794. doi:10.1007/s00421-014-3060-1

- Bell PG, Furber MJW, Van Someren K, Anton-Solanas A, Swart J. The Physiological Profile of a Multiple Tour de France Winning Cyclist ´. *Med Sci Sports Exerc*. 2016;49(1):115-123. doi:10.1249/MSS.00000000001068
- 18. Lamberts RP, Lambert MI, Swart J, Noakes TD. Allometric scaling of peak power output accurately predicts time trial performance and maximal oxygen consumption in trained cyclists. *Br J Sports Med.* 2012;46(1):36-41. doi:10.1136/bjsm.2010.083071
- 19. Wharton G, Steeds R, Allen J, et al. A minimum dataset for a standard adult transthoracic echocardiogram: a guideline protocol from the British Society of Echocardiography. *Echo Res Pract*. 2015;2(1):G9-G24. doi:10.1530/ERP-14-0079
- 20. Aune E, Baekkevar M, Roislien J, Rodevand O, Otterstad JE. Normal reference ranges for left and right atrial volume indexes and ejection fractions obtained with real-time three-dimensional echocardiography. 2009;i:738-744. doi:10.1093/ejechocard/jep054
- 21. Daanen HAM, Racinais S, Périard JD. Heat Acclimation Decay and Re-Induction: A Systematic Review and Meta-Analysis. *Sport Med.* 2018;48(2):409-430. doi:10.1007/s40279-017-0808-x
- Fox RH, Goldsmith R, Hunt TJ, Hampton IFG, Hunt TJ. Heat acclimatization by controlled hyperthermia in hot-dry and hot-wet climates. *J Appl Physiol*. 1967;22(1):39-46. doi:10.1152/jappl.1967.22.1.39
- 23. Sawka MN, Noakes TD. Does dehydration impair exercise performance? *Med Sci Sports Exerc*. 2007;39(8):1209-1217. doi:10.1249/mss.0b013e318124a664
- 24. Dill DB, Costill DL. Calculation of percentage changes in volumes of blood, plasma, and red cells in dehydration. *J Appl Physiol*. 1974;37(2):247-248. doi:10.1152/jappl.1974.37.2.247
- 25. Toner MM, Drolet LL, Pandolf KB. Perceptual and physiological responses during exercise in cool and cold water. *Percept Mot Skills*. 1986;62(1):211-220. doi:10.2466/pms.1986.62.1.211
- 26. Zhang H, Huizenga C, Arens E, Wang D. Thermal sensation and comfort in transient non-uniform thermal environments. *Eur J Appl Physiol*. 2004;92(6):728-733. doi:10.1007/s00421-004-1137-y
- 27. Moran DS, Shitzer A, Pandolf KB. A physiological strain index to evaluate heat stress. *Am J Physiol*. 1998;275(1):R129-34. doi:10.1152/ajpregu.1998.275.1.R129
- 28. Nerbass FB, Pecoits-Filho R, Clark WF, Sontrop JM, McIntyre CW, Moist L. Occupational Heat Stress and Kidney Health: From Farms to Factories. *Kidney Int Reports*. 2017;2(6):998-1008. doi:10.1016/j.ekir.2017.08.012
- 29. Ramanathan N. A new weighting system for mean surface temperature of the human body. *J Appl Physiol*. 1964;19:531-533. doi:10.1152/jappl.1964.19.3.531
- 30. Batterham AM, Hopkins WG. Making meaningful inferences about magnitudes. *Int J Sports Physiol Perform*. 2006;1(1):50-57.
- 31. Nielsen B, Hales J, Strange S, Christensen NJ, Warberg J, Saltin B. Human circulatory and thermoregulatory adaptations with heat acclimation and exercise in a hot, dry

environment. J Physiol. 1993;460:467-485.

- 32. Wolf MB. Hemoglobin-Dilution Method: Effect of Measurement Errors on Vascular Volume Estimation. *Comput Math Methods Med*. 2017;2017:3420590. doi:10.1155/2017/3420590
- 33. Keiser S, Meinild-Lundby AK, Steiner T, et al. Detection of blood volumes and haemoglobin mass by means of CO re-breathing and indocyanine green and sodium fluorescein injections. *Scand J Clin Lab Invest*. 2017;77(3):164-174. doi:10.1080/00365513.2016.1271908
- 34. Ertl A, Diedrich A, Raj SR. Techniques Used for the Determination of Blood Volume. *Am J Med Sci.* 2007;334(1):32-36. doi:10.1159/000367894
- 35. Burge CM, Skinner SL. Determination of hemoglobin mass and blood volume with CO: evaluation and application of a method. *J Appl Physiol*. 1995;79(2):623-631. doi:10.1152/jappl.1995.79.2.623
- 36. Garrett A, Rehrer N, Patterson M. Induction of heat acclimation in moderately and highly trained athletes. *Sport Med*. 2011;41(9):757-771.
- 37. Rodrigues ACT, de Melo Costa J, Alves GB, et al. Left ventricular function after exercise training in young men. *Am J Cardiol*. 2006;97(7):1089-1092. doi:10.1016/j.amjcard.2005.10.055
- 38. Nagueh SF, Smiseth OA, Appleton CP, et al. Recommendations for the Evaluation of Left Ventricular Diastolic Function by Echocardiography: An Update from the American Society of Echocardiography and the European Association of Cardiovascular Imaging. J Am Soc Echocardiogr. 2016;29(4):277-314. doi:10.1016/j.echo.2016.01.011
- 39. Neilan TG, Yoerger DM, Douglas PS, et al. Persistent and reversible cardiac dysfunction among amateur marathon runners. *Eur Heart J*. 2006;27(9):1079-1084. doi:10.1093/eurheartj/ehi813
- 40. Armstrong LE, Maresh CM. The Induction and Decay of Heat Acclimatisation in Trained Athletes. *Sport Med.* 1991;12(5):302-312. doi:10.2165/00007256-199112050-00003
- 41. Pelà G, Regolisti G, Coghi P, et al. Effects of the reduction of preload on left and right ventricular myocardial velocities analyzed by Doppler tissue echocardiography in healthy subjects. *Eur J Echocardiogr*. 2004;5(4):262-271. doi:10.1016/j.euje.2003.10.001
- 42. Nagueh SF, Middleton KJ, Kopelen HA, Zoghbi WA, Quinones MA. Doppler Tissue Imaging : A Noninvasive Technique for Evaluation of Left Ventricular Relaxation and Estimation of Filling Pressures. *JACC*. 1997;30(6):1527-1533. doi:10.1016/S0735-1097(97)00344-6
- 43. Dokainish H, Zoghbi WA, Lakkis N, et al. Optimal noninvasive assessment of Left Ventricular Filling Pressures: A comparison of Tissue Doppler Echocardiography and B-type natriuretic peptide in patients with Pulmonary Artery Catheters. *Circulation*. 2004;109:2432-2439. doi:10.1161/01.CIR.0000127882.58426.7A
- 44. Hellsten Y, Nyberg M. Cardiovascular Adaptations to Exercise Training. *Compr Physiol*. 2016;6(January):1-32. doi:10.1002/cphy.c140080

- 45. Wilson MG, Périard JD, Adamuz C, Farooq A, Watt V, Racinais S. Does passive heat acclimation impact the athlete's heart continuum? *Eur J Prev Cardiol*. 2019:DOI: 10.1177/2047487319836522 journals.sagepub.com. doi:10.1177/2047487319836522
- 46. Hare JL, Brown JK, Marwick TH. Performance of Conventional Echocardiographic Parameters and Myocardial Measurements in the Sequential Evaluation of Left Ventricular Function. *Am J Cardiol*. 2008;101(5):706-711. doi:10.1016/j.amjcard.2007.10.037
- Palmieri V, Arezzi E, Sabatella M, Celentano A. Interstudy reproducibility of parameters of left ventricular diastolic function: a Doppler echocardiography study. J Am Soc Echocardiogr. 2003;16(11):1128-1135. doi:https://doi.org/10.1067/j.echo.2003.07.003
- 48. Bain AR, Smith KJ, Lewis NC, et al. Regional changes in brain blood flow during severe passive hyperthermia: effects of PaCO2 and extracranial blood flow. *J Appl Physiol*. 2013;115:653-659.
- 49. Hellon R, Jones R, MacPherson R, Weiner J. Natural and Artificial Acclimatization to Hot Environments. *J Physiol*. 1956;132:559-576.
- Charlot K, Tardo-Dino P-EE, Buchet J-FF, et al. Short-term, low-volume training improves heat acclimatization in an operational context. *Front Physiol*. 2017;8(JUN):1-12. doi:10.3389/fphys.2017.00419
- 51. Périard JD, Racinais S, Sawka MN. Adaptations and mechanisms of human heat acclimation: Applications for competitive athletes and sports. *Scand J Med Sci Sport*. 2015;25(S1):20-38. doi:10.1111/sms.12408
- 52. Racinais S, Alonso JM, Coutts AJ, et al. Consensus recommendations on training and competing in the heat. *Br J Sports Med*. 2015;49(18):1164-1173. doi:10.1136/bjsports-2015-094915
- 53. Harrison MH. Effects on thermal stress and exercise on blood volume in humans. *Physiol Rev.* 1985;65(1):149-209. doi:10.1152/physrev.1985.65.1.149
- 54. Parsons IT, Stacey MJ, Woods DR. Heat Adaptation in Military Personnel : Mitigating Risk , Maximizing Performance. *Front Physiol*. 2019;10(December). doi:10.3389/fphys.2019.01485



Figure 1: The change in significant (p<0.05) echocardiographic parameters from pre exposure to post exposure in the HOT (heat acclimation) and TEMP (temperate exercise) groups. LV; left ventricle, IVC; inferior vena cava.

		TEMP		НОТ			
		Mean (SD)		Mean (SD)		Δ Cohen's D	P value
		Pre	Post	Pre	Post		
LV volumes							
EDV	(ml)	154 (13)	150 (15)	152 (21)	161 (16)	1.65 (0.39-2.91)	0.03*
ESV	(ml)	63 (4)	62 (6)	62 (12)	66 (12)	0.98 (-0.18-2.13)	0.17
SV (LVEDV-LVESV)	(ml)	91 (10)	88 (12)	90 (13)	96 (13)	1.29 (0.09-2.49)	0.05
SV (LVOT area x LVOT VTI)	(ml)	108 (16)	103 (9)	102 (9)	113 (8)	1.69 (0.42-2.96)	0.01*
Ejection fraction	(%)	59 (2)	58 (4)	60 (5)	59 (5)	0.17 (-0.92-1.27)	0.77
LV longitudinal function							
MAPSE (lateral)	(cm)	1.9 (0.4)	1.8 (0.2)	1.7 (0.3)	1.8 (0.3)	-0.06 (-1.19-1.07)	0.56
MAPSE (medial)	(cm)	1.4 (0.1)	1.5 (0.4)	1.5 (0.2)	1.4 (0.1)	0.11 (-1.02-1.24)	0.86
LV S'	(cm/s)	14 (2)	12 (1)	12 (2)	12 (2)	0.77 (-0.45-2.00)	0.97
RV longitudinal function							
TAPSE	(cm)	2.5 (0.3)	2.6 (0.3)	2.4 (0.2)	2.3 (0.3)	-0.75 (-1.88- 0.37)	0.17
RV S'	(cm)	16 (3)	13 (2)	15 (2.2)	13 (2.0)	0.41 (-1.71- 2.53)	0.50
Diastology							
E	(cm/s)	83 (20)	74 (8)	78 (13)	83 (6)	1.12 (-0.14-2.38)	0.10
А	(cm/s)	39 (12)	35 (7)	44 (14)	37 (7)	-0.29 (-1.48- 0.9)	0.11
E/A		2.3 (0.9)	2.2 (0.6)	1.8 (0.4)	2.3 (0.6)	1.08 (-0.18-2.33)	0.11
Ε'	(cm/s)	23 (4)	16 (4)	19 (2)	16 (4)	1.40 (0.10-2.71)	0.04*
Α'	(cm/s)	10 (2)	8 (3)	10 (3)	9 (2)	0.65 (-0.57 -1.86)	0.31
Atrial Volumes							
LA volume	(ml)	57 (10)	53 (11)	58 (20)	67 (19)	1.91 (0.6-3.23)	0.005**
RA Volume	(ml)	60 (17)	60 (19)	68 (16)	76 (19)	0.57 (-0.54-1.68)	0.33
Inferior Vena Cava							
IVC diameter expiration	(cm)	2.0 (0.3)	1.7 (0.5)	1.8 (0.4)	2.5 (0.6)	1.60 (0.26-2.94)	0.03*
IVC diameter inspiration	(cm)	0.9 (0.2)	0.6 (0.4)	0.9 (0.3)	1.2 (0.3)	1.98 (0.6-3.36)	0.03*